

Full Length Research Paper

Screening of *Trichoderma* species for virulence efficacy on seven most predominant phytopathogens

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In vitro studies on the efficacy of *Trichoderma* species against phytopathogens revealed the antagonistic potential of eight different species of *Trichoderma* isolated from the rhizosphere soils of varied locations of Uttar Pradesh and were evaluated *in vitro* against the most widely occurring soil inhabiting plant pathogens viz., *Fusarium oxysporum* f. sp. *ciceri*, *Alternaria solani*, *Phytophthora infestans*, *Pythium aphanidermatum*, *Sclerotium rolfsii*, *Bipolaris sorokiniana* and, *Rhizoctonia solani* and identify the most potential and effective strains *Trichoderma* with high antagonistic activity. Eight species of *Trichoderma* highly inhibited the growth of the seven test phytopathogens by producing volatile compounds showing variability in antagonistic potential of different *Trichoderma* spp. against the different pathogens tested. The test antagonists grow faster than the pathogen and produced inhibition zones. These antagonistic interactions influence the incidence and severity of the disease caused by the pathogen. The present communication describes the impact of different *Trichoderma* spp. on growth inhibition of plant pathogens under *in vitro* conditions. The data revealed that *Trichoderma reesei* (Tr(CSAU)) showed the maximum inhibition percentage of mycelial growth was recorded as against *Sclerotium rolfsii* (69.14%) followed by *Bipolaris sorokiniana* (80.33%), *Alternaria brasicae* (83.3%), while, in case of remaining plant pathogens such as *Pythium aphanidermatum*, *Fusarium oxysporum* f. sp. *ciceri*, *Trichoderma harzianum* (*Th.azad*) were reduced the highest radial mycelium growth of *Pythium aphanidermatum*, *Fusarium oxysporum* f. sp. *ciceri* (85 and 80%) respectively. Apart from *Trichoderma reesei* and *Trichoderma harzianum* (*Th.azad*), *Trichoderma viride* (01PP) also inhibited the mycelial growth of *Phytophthora infestans* (80.83%) and *Rhizoctonia solani* (70.42%).

Key words: Antagonism, *Trichoderma*, phytopathogens.

INTRODUCTION

Plants are a major source of food, fodder, medicines and many other useful products for humans. Diseases are the important biotic causes for low crop yield and poor quality seed. Pathogens being soil and seed borne, possess a great problem in disease management. Soil borne diseases are difficult to control and seed treatment with

fungicides does not protect the crop for longer periods. Continuous use of the same fungicide against the same pathogen results in the development of fungicide resistant strains of the pathogen (Shanmugam and Varma, 1998; Kumar and Dubey, 2001; Mamgain et al., 2013). Moreover, chemical measures may establish imbalances in the

microbiological community for the activity of beneficial organisms which otherwise improve the crop health. The demand for alternative to chemical control of plant pathogens has become stronger owing to the concerns about the safety and environmental aspects of chemicals. However, biological control offers the chance to improve crop production within the existing resources, besides avoiding the problem of pesticide resistance (Dekker, 1976; Khan et al., 2014). The genus *Trichoderma* is common filamentous imperfect fungi (*Deuteromycetes*), the most common saprophyte in the rhizosphere and found in almost all soils. Characterization for the antagonistic potential of *Trichoderma* spp. is the first step in utilizing the full potential of *Trichoderma* spp. for specific applications. *In vitro* screening of different pathogens is an effective and rapid method for identifying species with antagonistic potential. *Trichoderma*, a filamentous soil borne mycoparasitic fungus, has been shown to be effective against many soil borne plant pathogens (Papavizas, 1985; Pan et al., 2001; Jash and Pan, 2004) as they have more than one mechanism of action. Therefore, the study was conducted to evaluate the antagonistic activity of eight different *Trichoderma* species viz., *Trichoderma viride*, *T. harzianum*, *T. reesei*, *T. atroviride*, *T. asperellum*, *T. koningii*, *T. longibrachiatum* and *T. virens*, in inhibiting the growth of some most widely occurring soil inhabiting plant pathogens viz., *Fusarium oxysporum* f. sp. *ciceri*, *Alternaria solani*, *Pythium aphanidermatum*, *Phytophthora infestans*, *Sclerotium rolfsii*, *Bipolaris sorokiyana*, *Rhizoctonia solani* and identify *Trichoderma* spp. with a high antagonistic potential. Biological control of plant pathogens by microorganisms has been considered a more natural and environmentally acceptable alternative to the existing chemical treatment methods. *Trichoderma* spp. is now the most common fungal biological control agents that have been comprehensively researched and deployed throughout the world. Several fungal cell wall degrading enzymes, amongst them chitinase and glucanase, which seem to play an important role in the antagonistic action of *Trichoderma* against a wide range of fungal plant pathogens. The present study aimed to find out the efficiency of *Trichoderma* spp. against some phytopathogens.

MATERIALS AND METHODS

Isolation and purification of *Trichoderma* species

The isolation of eight *Trichoderma* spp. from rhizospheric zone of soil through serial dilution plate techniques (Johnson and Curl, 1972) on modified *Trichoderma* Selective Medium (TSM) (Saha and Pan, 1997). The green coloured colonies were identified by slide

culture technique and compared with taxonomic key of Rifai (1969) at genus and species level and deposit to ITCC division of plant pathology New Delhi for reconfirmation. After confirmation cultures of *Trichoderma* spp. were maintained on PDA slants and stored in the refrigerator at 4°C for further studies.

Isolation of plant pathogens

The pathogens were isolated from disease plants showing symptoms of disease. These isolated pathogens were identified, purified and tested for pathogenicity (Tapwal et al., 2011). The hyperparasitic potential of eight *Trichoderma* species were screened *in vitro* against seven test plant pathogens viz., *Fusarium oxysporum* f. sp. *ciceri*, *Alternaria solani*, *Rhizoctonia solani*, *Pythium aphanidermatum*, *Phytophthora infestans*, *Sclerotium rolfsii* and *Bipolaris sorokiyana* by dual culture plate technique and production of volatile and non-volatile metabolite part of Petri plate with respective test pathogen on the upper lid of plate served as control. Three replicates were maintained for each treatment. The assembly was opened after 72 h and the observations were recorded by measuring colony diameter of the test pathogen (in mm) in each plate and that of the control plates.

Efficacy of *Trichoderma* spp. on growth of the pathogens by dual-culture plate method

For testing antagonism in dual culture method (Morton and Stroube, 1955), a mycelial disc (6 mm) was cut from the margins of actively growing region of seven day old cultures of *Trichoderma* spp. and inoculated at one end of the petriplates (1 cm away from the edge of the plate) with sterilized potato dextrose agar (PDA) medium and simultaneously at the opposite end of a mycelial disc (6 mm) of the test pathogens. The experiments were conducted with three replications/plates for each treatment, while control plates were inoculated only by tested fungus. Plates were then incubated at 27 ± 1°C. Observation were recorded after seven days of inoculation including area covered by the *Trichoderma* spp. (eight *Trichoderma* spp.) and the pathogen while percent of inhibition was calculated using the following formula (Vincent, 1947):

$$\text{Percent growth of inhibition} = \frac{\text{colony growth in control plate} - \text{colony growth in intersecting plates}}{\text{colony growth in control}} \times 100$$

RESULTS

Our results explain that significant success in biocontrol is achieved under *in vitro* conditions. It is evident from the data presented in Table 1 and showed in Figures 1, 2 and 3, that the *Trichoderma* spp. suppressed the radial growth of different phytopathogens significantly on potato dextrose agar medium in the dual culture. *Trichoderma* spp. isolated from the rhizosphere soils of different location of Uttar Pradesh identified and confirmed on the basis of morphological and physiological characterization and micrometry observations revealed that they belong to eight different species viz., *Trichoderma viride*,

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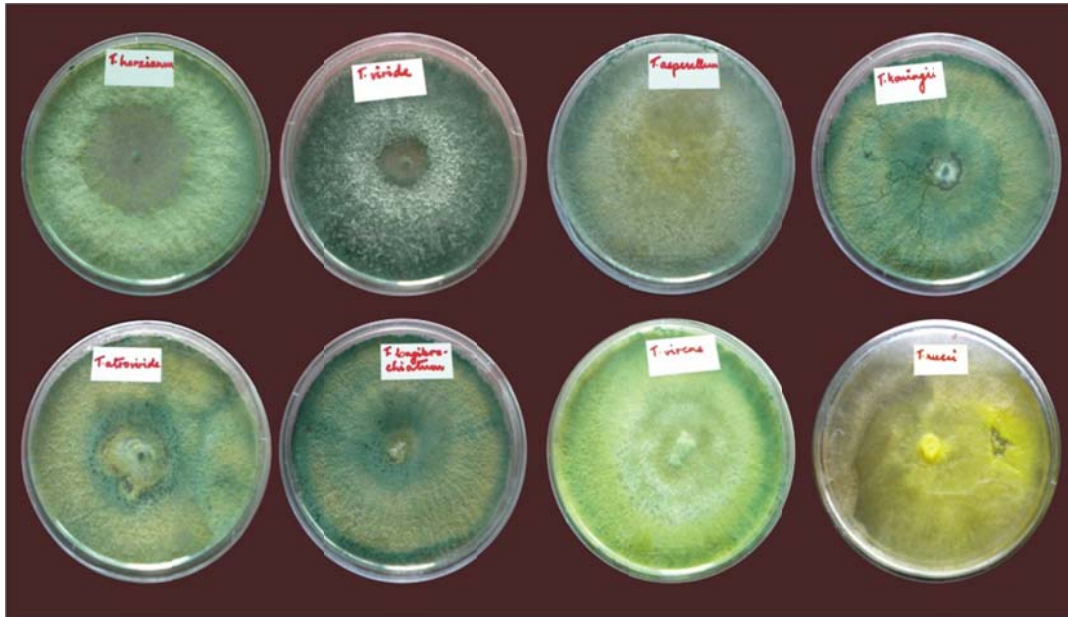


Figure 1. Eight *Trichoderma* spp. isolated from different locations of Uttar Pradesh.



Figure 2. Seven predominant plant pathogens.

T. harzianum, *T. reesei*, *T. atroviride*, *T. koningii*, *T. asperillum*, *T. virens*, *T. longibrachiatum* and *T. viride* was the most predominant species. The morphological and physiological characterization of these antagonistic species

was accomplished on the basis of colony color, growth rate, texture, growth patterns, size of phialides and phialospores.

Eight *Trichoderma* spp. were tested against most

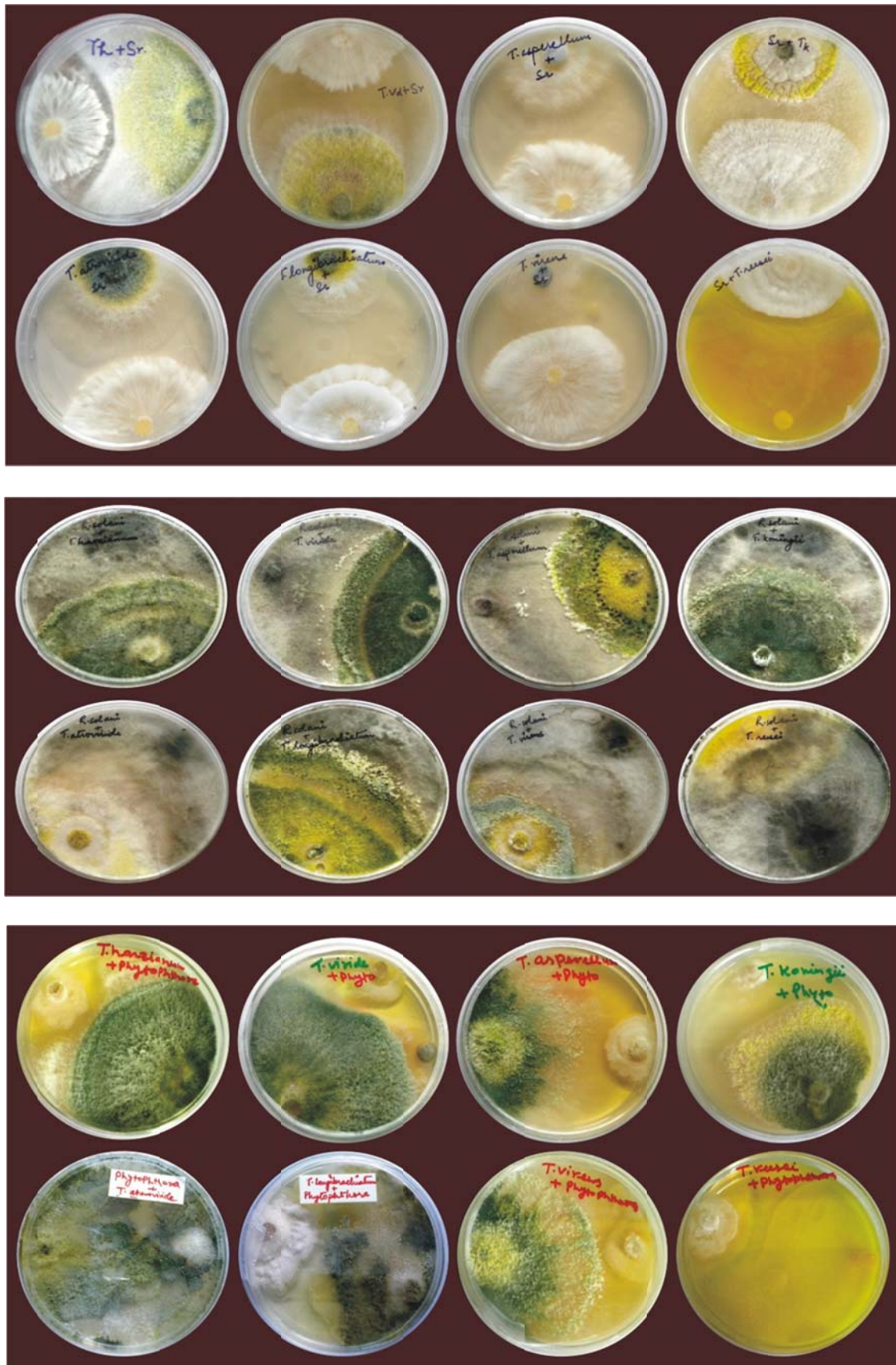


Figure 3. Confrontation test of *Trichoderma* spp. against notable plant pathogens.

Table 1. Submission of the gene sequences at NCBI database

| Strain No. | Name of Bioagent | Strain code | ITCC Acc. No | NCBI GenBank Accession No. | NBAIM, Mau | Source | GPS Location |
|------------|---------------------------|------------------------|--------------|----------------------------|------------|------------------|---|
| T1 | <i>T. harzianum</i> | Th azad | 6796 | KC800922 | TF-1271 | CSA Kanpur Nagar | Latitude: 25° 8' 34.821" Longitude: 81° 59' 2.979" |
| T2 | <i>T. viride</i> | 01PP | 8315 | JX119211 | TF-1272 | Hardoi | Latitude: 27° 23' 40.729" Longitude: 80° 7' 47.751" |
| T3 | <i>T. asperellum</i> | T _{asp} /CSAU | 8940 | KC800921 | TF-1270 | CSA Kanpur Nagar | Latitude: 25° 8' 34.821" Longitude: 81° 59' 2.979" |
| T4 | <i>T. koningii</i> | T _k (CSAU) | 5201 | KC800923 | TF-1269 | CSA Kanpur Nagar | Latitude: 26° 29' 33.384" Longitude: 80° 18' 6.518" |
| T5 | <i>T. atroviride</i> | 71 L | 7445 | KC 008065 | TF-1268 | Hardoi | Latitude: 26° 29' 28.323" Longitude: 80° 18' 26.361" |
| T6 | <i>T. longibrachiatum</i> | 21 PP | 7437 | JX978542 | TF-1267 | Kaushambi | Latitude: 26° 34' 27.61" Longitude: 79° 18' 24.623" |
| T7 | <i>T. virens</i> | T _{vi} (CSAU) | 4177 | KC800924 | TF-1266 | CSA Kanpur Nagar | Latitude: 25° 21' 39.794" Longitude: 81° 24' 11.414" |
| T8 | <i>T. reesei</i> | T _r (CSAU) | 8372 | KM999966 | TF-1273 | CSA farm | Latitude: 25° 21' 39.794" Longitude: 81° 24' 11.414" |

predominant seven phytopathogens in dual culture plates such as *Scelotium rolfsii*, *Alternaria brasicae*, *Pythium aphanidermatum*, *Phytophthora infestans*, *Alternaria brasicae*, *Bipolaris sorokiniana* and *Fusarium oxysporum* f. sp. *cicero* (Table 2). Among the eight *Trichoderma* spp. *Trichoderma reesei* (Tr(CSAU)) shows that maximum inhibition percentage of mycelial growth of pathogen was recorded (69.14%) against *Scelotium rolfsii* and minimum inhibition percentage was recorded (42.85%) by *Trichoderma atroviride* (71L), Similarly *Trichoderma viride* (01PP) found that maximum inhibition percentage of mycelial growth of pathogen was recorded (70.42%) against *Rhizoctonia solani* and minimum inhibition percentage was recorded (56.25%) by *Trichoderma atroviride* (71L), *Trichoderma harzianum* (Th. azad) revealed that maximum inhibition percentage of mycelial growth

of pathogen was recorded (85.00%) against *Pythium aphanidermatum* and minimum inhibition percentage was observed (55.55%) by *Trichoderma reesei* (Tr(CSAU)), *Trichoderma viride* (01PP) revealed that maximum inhibition percentage of mycelial growth of pathogen was recorded (80.83%) against *Phytophthora infestans* and minimum inhibition percentage was observed (66.25%) by *Trichoderma koningii* (T_k(CSAU)), *Trichoderma reesei* (Tr(CSAU)) revealed that maximum inhibition percentage of mycelial growth of pathogen was recorded (83.33%) against *Alternaria brasicae* and minimum inhibition percentage was observed (70.37%) by *Trichoderma atroviride* (71L), *Trichoderma asperellum* (T_{asp}/CSAU) revealed that maximum inhibition percentage of mycelial growth of pathogen was recorded (73.00%) against *Bipolaris sorokiniana* and minimum

inhibition percentage was observed (65.00%) by *Trichoderma atroviride* (71L), *Trichoderma harzianum* (Th. azad) revealed that maximum inhibition percentage of mycelial growth of pathogen was recorded (80.00%) against *Fusarium oxysporum* f.sp. *cicero* and minimum inhibition percentage was observed (60.00%) by *Trichoderma reesei* (Tr(CSAU)). From the above facts it was concluded that *Trichoderma harzianum* (Th. azad) shows the maximum inhibition against phytopathogens; *Pythium aphanidermatum* and *Fusarium oxysporum* f.sp. *cicero*, bioagents *Trichoderma viride* (01PP) shows the maximum inhibition against *Rhizoctonia solani* and *Phytophthora infestans*, *Trichoderma atroviride* (71 L) shows the minimum inhibition against four phytopathogens namely; *Scelotium rolfsii*, *Rhizoctonia solani*, *Alternaria brasicae* and

Table 2. Antagonistic activity of *Trichoderma* spp. against seven *phytopathogens* by dual culture method.

| Name of Bioagent | Culture No. | Source/ District | Id. No. | Average growth (mm) | % inhibition growth (mm) | Average growth (mm) | % inhibition growth (mm) | Average growth (mm) | % inhibition growth (mm) | Average growth (mm) | % inhibition growth (mm) | Average growth (mm) | % inhibition growth (mm) | Average growth (mm) | % inhibition growth (mm) | Average growth (mm) | % inhibition growth (mm) |
|---------------------------|------------------------|------------------------|---------|--------------------------|--------------------------------------|-------------------------------|-------------------------------|----------------------------|------------------------------|---|--------------------------|---------------------|--------------------------|---------------------|--------------------------|---------------------|--------------------------|
| | | | | <i>Sclerotium rolsii</i> | <i>Rhizoctonia solani</i> (Soyabean) | <i>Pythium aphanidermatum</i> | <i>Phytophthora infestans</i> | <i>Alternaria brasicae</i> | <i>Bipolaris sorokiniana</i> | <i>Fusarium oxysporum f.sp. ciceri</i> (CSAU) | | | | | | | |
| <i>T. harzianum</i> | Th azad | CSA Kanpur Nagar | 6796 | 34.68 | 50.48 | 25.00 | 68.75 | 13.33 | 85.00 | 21.60 | 72.92 | 16.66 | 81.48 | 26.00 | 65.50 | 45.00 | 80.00 |
| <i>T. viride</i> | 01PP | Hardoi | 8315 | 35.00 | 50.00 | 23.66 | 70.42 | 19.00 | 78.88 | 15.33 | 80.83 | 20.00 | 77.70 | 23.00 | 71.25 | 54.00 | 76.00 |
| <i>T. asperellum</i> | T _{asp} /CSAU | CSA Kanpur Nagar | 8940 | 33.30 | 52.38 | 28.33 | 64.58 | 25.66 | 71.48 | 25.00 | 68.75 | 25.00 | 72.22 | 21.00 | 73.00 | 78.00 | 65.30 |
| <i>T. koningii</i> | T _K (CSAU) | CSA Kanpur Nagar | 5201 | 36.66 | 47.62 | 27.66 | 65.42 | 21.00 | 76.66 | 27.00 | 66.25 | 21.66 | 75.93 | 24.60 | 69.25 | 54.00 | 76.00 |
| <i>T. atroviride</i> | 71 L | Hardoi | 7445 | 40.00 | 42.85 | 35.00 | 56.25 | 25.00 | 72.22 | 16.60 | 79.17 | 26.66 | 70.37 | 28.00 | 65.00 | 66.00 | 70.60 |
| <i>T. longibrachiatum</i> | 21 PP | Kaushambi | 7437 | 33.33 | 52.38 | 28.00 | 65.00 | 15.66 | 82.60 | 20.0 | 75.00 | 20.50 | 77.22 | 27.60 | 65.50 | 75.00 | 66.60 |
| <i>T. virens</i> | T _{vi} (CSAU) | CSA Kanpur Nagar | 4177 | 33.33 | 52.38 | 31.00 | 61.25 | 17.33 | 80.74 | 23.00 | 71.25 | 18.33 | 70.63 | 23.30 | 70.80 | 60.00 | 73.30 |
| <i>T. reesei</i> | Tr(CSAU) | CSA Kanpur Nagar | 7284 | 21.66 | 69.14 | 26.66 | 66.66 | 40.00 | 55.55 | 15.00 | 80.33 | 15.00 | 83.33 | 22.00 | 72.50 | 90.00 | 60.00 |

Bipolaris sorokiniana and finally, *Trichoderma reesei* (Tr(CSAU)) shows the maximum inhibition against *Sclerotium rolsii*, *Alternaria brasicae* and minimum inhibition against *Pythium aphanidermatum*, *Fusarium oxysporum f.sp. ciceri*, respectively.

Experimental design and statistical analysis

Statistical analysis was performed following completely randomized block design (CRBD) with three replicates in each treatment.

DISCUSSION

In the study it may be concluded that among the eight different species of *Trichoderma* exhibited different growth inhibition percentage against the tested most predominant phytopathogens with variability in the antagonistic potential. *Trichoderma reesei*, *Trichoderma harzianum* (T.azad), and also *Trichoderma viride* (01PP) showed high antagonistic potential against tested phytopathogens. Plant pathogenic fungi are a widespread problem and the use of chemicals is hardly successful (Anand and Jayarama, 2009).

However, the high cost associated with the use of chemical (fungicides) to control disease caused by soil borne fungi is a limiting factor in the profitability of crop production, in this case biological control could be the best alternative. *Trichoderma* is the most commonly used fungal biological control agent and have long been known as effective antagonists against plant pathogenic fungi (Chet et al., 1981; Papavizas, 1985). Some of the species of *Trichoderma* included in the present study significantly inhibited several pathogens (Dubey, 2003).

All the *Trichoderma* spp. restricted the growth of

all the seven test phytopathogens in their own way. Thus, it is well known that all species isolated from different samples of soil are not equally antagonistic to phytopathogen and searching of effective and potential species to locally suit the purpose is important.

The results reported, suggests that the species of *T. harzianum*, *T. viride*, *T. reesei* and *T. atroviride* were more capable of influencing the growth of tested seven pathogens in dual culture. Similarly, isolates of different *Trichoderma* spp. to control soil borne phytopathogens have been reported to differ in their effectiveness (Rama and Krishna, 2000; Anand and Jayarama, 2009; Singh et al., 2013; Kumar et al., 2014). This result is a pioneer information that particular isolate from a particular location can be employed in bulk for treatment of disease incidence.

Conclusion

In our study it was concluded that *Trichoderma harzianum* (Th. azad) show the maximum inhibition against pytopathogens; *Pythium aphanidermatum* and *Fusarium oxysporum* f.sp. *ciceri*, bioagents *Trichoderma viride* (01PP) shows the maximum inhibition against *Rhizoctonia solani* and *Phytophthora infestans* while *Trichoderma atroviride* (71 L) shows the minimum inhibition against four pytopathogens namely; *Sclerotium rolfsii*, *Rhizoctonia solani*, *Alterneria brasicae* and *Bipolaris sorokiniana*. Finally, *Trichoderma reesei* (Tr(CSAU)) shows the maximum inihibiton against *Sclerotium rolfsii*, *Alterneria brasicae* and minimum inhibition against *Pythium aphanidermatum*, *Fusarium oxysporum* f.sp. *cicero*, respectively.

Conflict of interests

The authors did not declare any conflict of interest.

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REFERENCES

- Anand S, Jayarama R (2009). Biocontrol potential of *Trichoderma* Sp. against plant pathogens. Int. J. Agric. Sci. 2:30-39.
- Chet I, Harman GE, Baker R (1981). *Trichoderma hamatum* its hyphal interaction with *Rhizoctonia solani* and *Pythium* spp. Microb. Biol. 7 29-38.
- Dubey SC (2003). Integrated management of web blight of urd/mung bean by bio-seed treatment. Indian Phytopathol. 56:34-38.
- Jash S, Pan S (2004). Evaluation of mutant isolates of *T. harzianum* against *R. solani* causing seedling blight of green gram. Ind J. Agric. Sci. 74:190-193.
- Johnson LF, Curl EH (1972). Methods for research on the ecology of soil borne plant pathogens. Burgess Publ. Co., Minneapolis.
- Khan F, Mazid M, Khan TA, Patel HK, Roychowdhury R (2014). Plant derived pesticides in control of lepidopteran insects: dictum and directions. Res. J. Biol. 2: 1 - 10.
- Kumar D, Dubey SC (2001). Management of collar rot of pea by the integration of biological and chemical methods. Indian Phytopathol. 54:62-66.
- Kumar V, Shahid M, Srivastava M, Singh A, Pandey S, Sharma A, Srivastava YK (2014). Antagonistic effect of rhizospheric *Trichoderma* species against soil borne pathogens. Progress. Res. 9(special): 408-410
- Mamgain A, Roychowdhury R, Tah J (2013). *Alternaria* pathogenicity and its strategic controls. Res. J. Biol. 1: 1 - 9.
- Morton DT, Stroube WH (1955). Antagonistic and stimulatory effects of microorganism upon *Sclerotium rolfsii*. Phytopathology 45: 419-420.
- Pan S, Roy A, Hazra S (2001). *In vitro* variability of biocontrol potential among some isolates of *Gliocladium virens*. Adv. Plant Sci. 14: 301-303.
- Papavizas GC (1985). *Trichoderma* and *Gliocladium*: Biology, ecology and potential for biocontrol. Ann. Rev. Phytopathol. 23: 23-54.
- Rama BRM, Krishna MKVM (2000). Efficacy of *Trichoderma* spp. In the management of collar rot of groundnut caused by *Aspergillus niger*, Van Tieghem. Indian J. Plant Prot. 28. 197-199.
- Rifai MA (1969). A revision of the genus *Trichoderma*. Mycol. Pap. 116: 1-116.
- Saha DK, Pan S (1997). Quantitative evaluation of some specific media of *Trichoderma* and *Gliocladium* spp. J. Mycopathol. Res. 35: 7-13.
- Shanmugam V, Varma AS (1998). *In vitro* evaluation of certain fungicides against selected antagonists of *Pythium aphanidermatum*, the incitant of rhizome rot of ginger. Abs J. Mycol. Plant Pathol. 28: 70.
- Singh A, Shahid M, Srivastava M, Kumar V, Bansal A (2013). Antagonistic activity of *Trichoderma viride* against different pathogens of *Fusarium oxysporum* isolated from legume crops of UP. Progress. Res. 8(1):47-50.
- Tapwal A, Singh U, da Silva JAT, Singh G, Garg S, Kumar R (2011). *In vitro* antagonism of *Trichoderma viride* against Five Phytopathogens. Pest Technol. 5(1):59-62.