



Effectiveness of Three Fruit Seed Extracts as Larvicide against Three Major Mosquito Vectors *Aedes aegypti* Linnaeus, *Culex quinquefasciatus* Say and *Anopheles gambiae* Giles (Diptera: Culicidae)

Lame Younoussa^{1*}, Kary Mallam Oumarou², Theodora Kopa Kowa³,
Serge Eteme Enama¹, Gabriel Agbor Agbor³ and Elias Nchiwan Nukenine⁴

¹Department of Animal Biology and Physiology, Faculty of Science, University of Yaounde 1,
P.O. Box 812, Yaounde, Cameroon.

²School of Veterinary Medicine and Sciences, University of Ngaoundere P.O. Box 454, Ngaoundere,
Cameroon.

³Centre for Research on Medicinal Plants and Traditional Medicine, Institute of Medical Research and
Medicinal Plants Studies, P.O. Box 13033, Yaounde, Cameroon.

⁴Department of Biological Sciences, Faculty of Science, University of Ngaoundere, P.O. Box 454,
Ngaoundere, Cameroon.

Authors' contributions

This work was carried out in collaboration among all authors. Authors LY, KMO and ENN designed the study. Author LY performed the statistical analysis. Authors LY and ENN wrote the protocol and wrote the first draft of the manuscript. Authors LY, TKK, SEE and GAA managed the analyses of the study. Authors LY and KMO managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJTDH/2020/v41i2330415

Editor(s):

(1) Dr. Cihad Dunder, Ondokuz Mayıs University, Turkey.

Reviewers:

(1) Anil Prakash, ICMR - National Institute for Research in Environmental Health, India.

(2) Nur Alvira Pascawati, Respati University of Yogyakarta, Indonesia.

Complete Peer review History: <http://www.sdiarticle4.com/review-history/64536>

Original Research Article

**Received 25 October 2020
Accepted 30 December 2020
Published 31 December 2020**

ABSTRACT

The CH₂Cl₂-MeOH (30:70 v/v) extracts of the seeds of *Mangifera indica* (Mango), *Persea americana* (Avocado) and *Dacryodes edulis* (African plum) were evaluated for potential mosquito larvicidal activity against 3rd and 4th instar larvae of *Aedes aegypti*, *Culex quinquefasciatus* and

*Corresponding author: Email: younoussalame@yahoo.com;

Anopheles gambiae. Extracts were diluted with 1 mL of methanol and concentrations ranging from 1000 to 125 mg/L in 4 replicates each, were prepared in the volume of 100 mL in the plastic cups (250 mL). A volume of 1 mL of methanol added to 99 mL of tap water was prepared as negative control and Bi-one (1000 mg/L) constituted a positive control. In each test solution, 25 larvae of each mosquito species were separately transferred and larval mortality was recorded after 24 h post-treatment. As results, the three plant seed extracts applied at 1000 mg/L caused for at least 79% mortality of each mosquito species larvae assessed. The seed extract of *P. americana* (LC₅₀ of 98.31, 129.24 and 136.26 mg/L, respectively against *An. gambiae*, *Ae. aegypti* and *Cx. quinquefasciatus* larvae) was the most potent followed by *D. edulis* (LC₅₀ of 176.87 mg/L for *An. gambiae*, 198.68 mg/L for *Ae. aegypti* and 201.70 mg/L for *Cx. quinquefasciatus*) and *M. indica* (LC₅₀ of 258.98 mg/L for *An. gambiae*, 297.35 mg/L for *Ae. aegypti* and 435.45 mg/L for *Cx. quinquefasciatus*). Globally, all the seed extracts were more toxic against *An. gambiae* larvae compared to other mosquito species and need further exploration for the development of a new botanical larvicide to reduce mosquito densities.

Keywords: Larvicidal; CH₂Cl₂-MeOH seed extracts; mosquito species; *Anopheles gambiae*.

1. INTRODUCTION

Mosquitoes are regarded as the most serious insect pests of public health importance due to their role in vectoring various pathogens such as protozoa, nematodes and arboviruses causing diseases to human and animals [1]. In African countries, mosquito-borne diseases like yellow fever, dengue fever, malaria and lymphatic filariasis are prevalent resulting in millions of deaths every year [2].

Lymphatic filariasis (LF), a neglected tropical disease and a serious health threat in Africa is principally caused by the filarial nematodes *Wuchereria bancrofti*, *Brugia timori* and *B. malayi*. Although species belonging to five mosquito genera i.e. *Ochlerotatus*, *Culex*, *Aedes*, *Anopheles* and *Mansonia*, are involved in the transmission of that filariasis, *Culex quinquefasciatus* is its principal vector in tropical and subtropical regions [3,4]. In 2019, an estimated 339.3 million people in 32 countries of WHO African Region were infected by the LF and required mass drug administration [5]. In Cameroon, Nana-Djeunga et al. [6] reported that LF infection is distributed nationwide.

Arboviruses causing yellow fever, dengue, chikungunya, infections mainly transmitted through the bites of tiger mosquito *Aedes aegypti*, are common in Africa. In Cameroon, increasing prevalence of dengue infection has been reported during the last decade [7,8,9,10,11]. In view of scarce availability of dengue vaccine, vector control remains the important tool to prevent and control this disease. Despite the availability of Yellow Fever (YF) vaccine, it remains a major public health problem

in Sub-Saharan Africa. Entomological studies conducted in Cameroon and Democratic Republic of Congo have implicated *Ae. aegypti* in the infection, transmission and dissemination of YF [12].

Malaria, transmitted through the bites of *Anopheles* spp, still remains the major public health problem particularly in sub-Saharan Africa. In 2018, WHO African region contributed to 94% of estimated malaria deaths globally of which 67% deaths occurred in children below 5 years [13]. In Cameroon, approximately 6,228,154 malaria cases and about 11,192 deaths were reported in 2018 [13].

Mosquito vector control options include chemical and biological control methods. The most commonly used control method involves application of synthetic chemical insecticides such as pyrethroids, organophosphates, organochlorines, pyrroles carbamates and phenyl pyrazole against mosquitoes [14]. This method, though effective, suffers from the disadvantages of the developing resistance in the target species against these chemicals and the persistence of their residue in the environment which is harmful to the human being and also non-target organisms [15,16]. This, calls for the search of alternative substitute for the chemical insecticides. Botanicals / plant products could be a potential alternative in this regard because of their target specificity, eco-friendly nature, biodegradability and cost-effectiveness. Besides, plant products contain a battery of phytochemicals that limits the development of resistance mosquito vectors [17]. Therefore, various parts such as leaf, bark, stem, flower, fruit, seed and root of the potential plants have

been investigated for their insecticidal properties against many insect pests. These plant parts may contain alkaloids, flavonoids, saponins, tannins, and phenolic compounds that possess insecticidal properties [18]. Thousands of tons of avocado, mango and African plum fruits are produced yearly and their seeds are generally discarded, thereby generating waste of environmental concern [19].

Persea americana Mill. (Lauraceae) commonly called avocado pear is a fruit tree native to Central America. This plant is widely cultivated in subtropical zones. Orange pigment from its seeds is used as a natural colorant [20,21]. The seed extracts are used in traditional medicine to manage hypertension and in the treatment of dysentery, diarrhea, intestinal parasites, skin infection and tooth ache [22,23]. Extracts have been reported to have larvicidal properties against *An. stephensi*, *Ae. aegypti*, *Cx. quinquefasciatus* mosquito larvae [24,25,26].

Dacryodes edulis (G. Don) H. J. Lam (Burseraceae), also known as safou or African plum, is an evergreen tree indigenous to and largely distributed in Gulf of Guinea and Central African regions [27]. As medicinal plant, it is used to treat leprosy, dysentery, anaemia, and also possesses antioxidant, antibacterial and antidiarrhoeal properties [28-30]. Aqueous, ethanol and hexane extracts of the leaves and seeds of *D. edulis* have been found to possess good larvicidal activity against the larvae of *Ae. vittatus*, *An. gambiae* and *Cx. quinquefasciatus* mosquito species [31,32].

Originated from South-East Asia, *Mangifera indica* L. (Anacardiaceae), the mango is a tree largely grown in tropical and subtropical regions for its fruits [33]. Tons of mango fruits are industrially processed or locally consumed for its pericarp and seeds are discarded and consequently generate a considerable amount of waste in the environment. Acetone, chloroform, and methanol leaf extracts of *M. indica* exhibited a moderate larvicidal activity against *Cx. quinquefasciatus* larvae [34]. *M. indica* crude extract caused substantial larvicidal activity against *Ae. aegypti* larvae [35].

The present study aimed to evaluate the larvicidal efficacy of the seed extract of three common edible fruits in Cameroon against the major sub-Saharan Africa mosquito vectors including *Ae. aegypti*, *Cx. quinquefasciatus* and *An. gambiae*.

2. MATERIALS AND METHODS

2.1 Plant Material Collection

The ripe Avocado, African plum and Mango fruits were purchased from Melen market in the Mfoundi Division, region of Centre, Cameroon in May, June and October 2019, respectively. Seeds of each plant were removed from the fruits, cleaned with tap water, chopped in small pieces and grinded in the electric blender. The paste, thus obtained was dried in the oven set at 60°C for 24 h to obtain a dry seed powder. Each plant seed powder were packaged in black plastics and stored in the refrigerator until needed for extraction.

2.2 Extraction of the Plant Seeds

The extraction of the plant seeds was carried out in the laboratory of Phytochemistry, Institute of Medical research and Medicinal Plant Studies of Yaounde in Cameroon. Each dried seed powder was separately soaked in the mixture of CH₂Cl₂-MeOH solvent in proportion (30:70 v/v) for 72 h in which it was manually stirred twice a day. After 3 days of maceration, the macerate was filtered using Whatman No. 1 filter paper. The filtrate was concentrated using rotary evaporator and each dried plant seed extract was kept in dark glass bottles in the refrigerator until their uses for phytochemical screening tests and mosquito larvae tests. For each plant seed powder, extraction yield was calculated using the following formula.

Extraction yield (%) =

$$\frac{\text{Weight of plant seed extract obtained (g)}}{\text{Weight of plant seed powder used (g)}} \times 100$$

2.3 Phytochemical Screening

Seeds of all the three plants were investigated for the presence of phytochemicals likely having insecticidal activity such as alkaloids, flavonoids, phenolic compounds, saponins, tannins, terpenoids and steroids following Harbone [36] in the laboratory of Phytochemistry, IMPM, Yaounde, Cameroon.

2.4 Mosquito Species Collection and Rearing

The larvae of the three mosquito species (*An. gambiae*, *Ae. aegypti* and *Cx. quinquefasciatus*) were collected from their natural breeding sites in the Yaounde town. In the laboratory, mosquito larvae were separated in sub-families wise

(Culicinae and Anophelinae) based on the morphological identification keys adopted by Azari-Hamidian and Harbach [37] and Gillies and Coetzee [38].

The larvae were fed with biscuits and crayfish (3:1) till they turn into pupae. After pupation species wise, pupae were transferred into 30 cm × 30 cm × 35 cm mosquito cages for emergence. Three days after emergence, mosquitoes were allowed to feed on a rabbit restrained in a wire cage. A bowl lined with filter paper containing tap water was provided in each cage for oviposition. Eggs of each mosquito species were transferred in to buckets containing well water for hatching. Third and fourth instar larvae of that 1st generation were used for the larvicidal test.

2.5 Larvicidal Bioassays

The standard procedure method described by WHO [39] was followed to carry out the larvicidal activity of the extract of the plant seeds against the 3rd and 4th instar larvae of *An. gambiae*, *Ae. aegypti* and *Cx. quinquefasciatus* in the laboratory. Indeed, plant extracts were previously diluted with 1 mL of an emulsifier Tween-40 and 4 different concentrations of 1000, 500, 250 and 125 mg/L were prepared in the total volume of 100 mL of tap water in the plastic cups (250 mL) and each concentration was repeated 4 times. The volume of 1 mL of methanol was added to 99 mL of tap water constituted a negative control while the commercial insecticide Bi-one (Dichlovos 49%) tested at the recommended concentration of 1000 mg/L was used as positive control. In each concentration solution test and controls (negative and positive controls), 25 larvae of each mosquito species were separately transferred and larval mortality was monitored

after 24 h post treatment. Is declared as dead, when mosquito larval did not longer move even pinched with an entomological needle. The mortality percentages were calculated and then corrected using Abbott [40] formula below if the mortality in the negative control is comprised between 5 and 20%.

$$\text{Mortality (\%)} = \frac{\text{Number of dead larvae in test/controls}}{\text{Total number of larvae used}} \times 100$$

$$\text{Corrected mortality (\%)} = \frac{(\text{Number of dead larvae in control} - \text{Number of dead larvae in test})}{(100 - \text{Number of dead larvae in control})} \times 100$$

2.6 Statistical Analyses

The corrected larval mortality was subjected to the Analysis of Variance (ANOVA) using the Statistical Package for the Social Science (SPSS version 16.0). For means comparison, Tukey test at P= 0.05 was deployed and Probit analysis [41] was performed to determine the concentrations that caused 50% (LC₅₀) and 95% (LC₉₅) mortality of mosquito larvae.

3. RESULTS

3.1 Extraction Yield

From *D. edulis* (360 g), *P. americana* (300 g) and *M. indica* (270 g) plant seed powders macerated in CH₂Cl₂-MeOH solvent (30:70 v/v), extraction yields of 9.41, 7.01 and 11.32%, respectively were obtained (Table 1). However, the extraction yield of *M. indica* (11.32%) was slightly high compared to *D. edulis* and *P. americana* yields.

Table 1. Phytochemical screening results of CH₂Cl₂-MeOH seed extracts

Phytochemical components	Plant species		
	<i>Dacryodes edulis</i>	<i>Persea americana</i>	<i>Mangifera indica</i>
Alkaloids	+++	+	+
Flavonoids	++	+	++
Phenolic compounds	+	++	-
Saponins	++	+++	+
Tannins	++	+	-
Steroids	-	+	-

-=absent; +=present at low concentration; ++=present at moderate concentration; +++= present at high concentration

3.2 Phytochemical Screening

High concentrations of alkaloids in *D. edulis*; that of saponins in *P. americana* and flavonoids in *M. indica* was detected in phytochemical screening of the plant seed extracts. In general, *M. indica* seed extract was relatively poor in various phytochemicals (Table 1).

Table 2 presents the mortality percent and LC₅₀ and LC₉₅ (mg/L) values of Plant seed extracts against larvae of *An. gambiae* 24 h post-treatment. Globally, the three plant seed extracts exhibited a significant (P<0.001) larvicidal activity and that efficacy augmented with the increasing concentration of the plant seed extracts. Treated with *P. americana* seed extract, the mortality of *An. gambiae* larvae ranged significantly (F_(5, 17) = 747.34; P<0.001) from 63% at 125 mg/L to 100% at 1000 mg/L. With the seed extract of *M. indica*, a moderate larval mortality of *An. gambiae* varying significantly (F_(5, 17) = 377.70; P<0.001) from 21% (at 125 mg/L) to 90% at 1000 mg/L) was recorded. Larval mortality of *An. gambiae* ranged significantly (F_(5, 17) = 545.76; P<0.001) from 39% (at 125 mg/L) to 100% (at 1000 mg/L) when tested with *D. edulis* seed extract.

After 24 h post-exposure of *An. gambiae* larvae to plant seed extracts, *P. americana* (LC₅₀ = 98.31 mg/L and LC₉₅ = 379.89 mg/L) was revealed as the most effective against *An. gambiae* larvae compared to *D. edulis* (LC₅₀ = 176.87 mg/L and LC₉₅ = 781.44 mg/L) and *M. indica* (LC₅₀ = 258.98 mg/L and LC₉₅ = 1338.45 mg/L).

Results of mortality percent, LC₅₀ and LC₉₅ values of *Ae. aegypti* larvae exposed for 24 h to the seed extracts of *P. americana*, *M. indica* and *D. edulis* are presented in Table 3. All the plant seed extracts assessed caused a significant larvicidal activity against *Ae. aegypti* larvae and that activity varied with the increasing concentration of each plant seed extracts tested. Tested at the lowest concentration of 125 mg/L, *P. americana* seed extract caused 53% mortality of *Ae. aegypti* and significantly (F_(5, 17) = 588.52; P<0.001) reached 100% mortality when applied at the highest dose of 1000 mg/L. The seed extract

of *M. indica* caused also a moderate mortality of *Ae. aegypti* larvae varying significantly (F_(5, 17) = 209.06; P<0.001) from 16% at the lowest dose (125 mg/L) to 85% at the highest dose (1000 mg/L). *D. edulis* seed extract exhibited also a high larvicidal activity against *Ae. aegypti* larvae and that efficacy ranged significantly (F_(5, 17) = 307.04; P<0.001) from 32% at 125 mg/L to 97% at 1000 mg/L.

As against *An. gambiae*, the seed extract of *P. americana* (LC₅₀ = 129.24 mg/L and LC₉₅ = 588.52 mg/L) was found to be the most effective against *Ae. aegypti* larvae compared to *D. edulis* (LC₅₀ = 198.68 mg/L and LC₉₅ = 977.79 mg/L) and *M. indica* (LC₅₀ = 297.34 mg/L and LC₉₅ = 1739.77 mg/L) seed extracts.

Table 4 presents the mortality rate of *Cx. quinquefasciatus* larvae exposed for 24 h to the seed extracts of *P. americana*, *M. indica* and *D. edulis*. In general, the three plant seed extracts tested showed a significant toxicity against the larvae of *Cx. quinquefasciatus* and that activity significantly increased with the increasing concentration of each plant seed extract.

The mortality of *Cx. quinquefasciatus* larvae ranged significantly (F_(5, 17) = 321.24; P<0.001) from 47% (at 125 mg/L) to 93% (at 1000 mg/L) when exposed to the seed extract of *P. americana*. After application of the seed extract of *M. indica*, mortality of *Cx. quinquefasciatus* larvae varied significantly (F_(5, 17) = 269.30; P<0.001) from 8% (at 125 mg/L) to 79% (at 1000 mg/L). Tested with the seed extract of *D. edulis*, the larval mortality of *Cx. quinquefasciatus* ranging significantly (F_(5, 17) = 539.17; P<0.001) from 33% (at 125 mg/L) to 90% (at 1000 mg/L) was registered after 24 h post-treatment.

Among the three plant seed extracts tested, the seed extract of *P. americana* (LC₅₀ = 136.26 mg/L and LC₉₅ = 1394.20 mg/L) was revealed as the most potent against *Cx. quinquefasciatus* larvae compared to *D. edulis* (LC₅₀ = 201.70 mg/L and LC₉₅ = 1393.46 mg/L) and *M. indica* (LC₅₀ = 435.45 mg/L and LC₉₅ = 2208.24 mg/L) seed extracts.3

Table 2. Mortality percentage of mosquito larvae and LC₅₀ as well as LC₉₅ (mg/L) values of *Persea americana*, *Mangifera indica* and *Dacryodes edulis* CH₂Cl₂-MeOH seed extracts after 24 h post-exposure against *Anopheles gambiae* larvae in the laboratory (26±3°C, 74±4% R.H.)

Plant species	Conc (mg/L)	% mortality	Slope±SE	R ²	LC ₅₀ (CI at 95%)	CL ₉₅ (CI at 95%)	χ ²
<i>Persea americana</i>	0	0.00±0.00d					
	125	63.00±2.51c					
	250	85.00±1.91b	2.80±0.20	0.62	98.31 (85.71-109.67)	379.89 (340.35-436.10)	17.33 ^{ns}
	500	98.00±1.15a					
	1000	100.0±0.00a					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F _(5,17)	747.34***					
<i>Mangifera indica</i>	0	0.00±0.00f					
	125	21.00±2.51e					
	250	52.00±2.82d	2.30±0.11	0.79	258.98 (240.59-277.71)	1338.45 (1159.43-1587.89)	17.88 ^{ns}
	500	75.00±02.51c					
	1000	90.00±1.15b					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F _(5,17)	377.70***					
<i>Dacryodes edulis</i>	0	0.00±0.00e					
	125	39.00±1.91d					
	250	61.00±3.00c	2.54±0.13	0.85	176.87 (154.10-198.72)	781.44 (646.25-1008.17)	30.88**
	500	84.00±1.63b					
	1000	100.0±0.00a					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F _(5,17)	545.76***					

Mean of mortality percent ± standard error within a column followed by the same letter did not differ significantly according to Tukey test at P= 0.05; ^{ns}P>0.05; **P<0.01; ***: p<0.001; CI= Confidence interval; SE= Standard error; R²=Coefficient of determination; χ² = Chi-square; Number of replicates: 4

Table 3. Mortality percentage of mosquito larvae and LC₅₀ as well as LC₉₅ (mg/L) values of *Persea americana*, *Mangifera indica* and *Dacryodes edulis* CH₂Cl₂-MeOH seed extracts after 24 h post-exposure against *Aedes aegypti* larvae in the laboratory (26±3°C, 74±4% R.H.)

Plant species	Conc (mg/L)	% mortality	Slope±SE	R ²	LC ₅₀ (CI)	CL ₉₅ (CI)	χ ²
<i>Persea americana</i>	0	0.00±0.00d					
	125	53.00±1.91c					
	250	70.00±2.58b	2.49±0.15	0.78	129.24	588.52	30.14**
	500	93.00±1.91a			(107.36-149.02)	(488.36-762.00)	
	1000	100.0±0.00a					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F(5, 17)	588.52***					
<i>Mangifera indica</i>	0	0.00±0.00f					
	125	16.00±1.63e					
	250	52.00±2.82d	2.14±0.11	0.75	297.34	1739.77	44.90***
	500	68.00±4.32c			(255.37-342.72)	(1289.66-2677.62)	
	1000	85.00±3.00b					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F(5, 17)	209.06***					
<i>Dacryodes edulis</i>	0	0.00±0.00e					
	125	32.00±2.30d					
	250	61.00±3.00c	2.37±0.12	0.82	198.68	977.79	32.42**
	500	79.00±3.00b			(172.61-224.20)	(792.20-1298.81)	
	1000	97.00±1.91a					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F(5, 17)	307.04***					

Mean of mortality percent ± standard error within a column followed by the same letter did not differ significantly according to tukey test at p= 0.05; **p<0.01; ***p<0.001; ci= confidence interval; se= standard error; r²=coefficient of determination; χ² = chi-square; number of replicates: 4

Table 4. Mortality percentage of mosquito larvae and LC₅₀ as well as LC₉₅ (mg/L) values of *Persea americana*, *Mangifera indica* and *Dacryodes edulis* CH₂Cl₂-MeOH seed extracts after 24 h post-exposure against *Culex quinquefasciatus* larvae in the laboratory (26±3°C, 74±4% R.H.)

Plant species	Conc (mg/L)	% mortality	Slope±SE	R ²	LC ₅₀ (CI)	CL ₉₅ (CI)	χ ²
<i>Persea americana</i>	0	0.00±0.00e					
	125	47.00±2.51d					
	250	68.00±2.82c	1.73±0.11	0.76	136.26	1394.20	22.13ns
	500	84.00±1.63b			(110.24-160.50)	(943.33-1711.29)	
	1000	93.00±2.51ab					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F(5, 17)	321.24***					
<i>Mangifera indica</i>	0	0.0±0.00e					
	125	8.00±01.63e					
	250	33.00±3.00d	2.33±0.11	0.88	435.45	2208.24	26.79*
	500	55.00±2.51c			(392.99-484.95)	(1736.65-3024.92)	
	1000	79.00±3.41b					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F(5, 17)	269.30***					
<i>Dacryodes edulis</i>	0	0.00±0.00f					
	125	33.00±1.91e					
	250	58.00±2.00d	1.96±0.11	0.78	201.70	1393.46	11.85ns
	500	80.00±2.30c			(182.94-220.16)	(1174.40-1717.72)	
	1000	90.00±1.15b					
	Bi-one (1000 mg/L)	100.0±0.00a					
	F(5, 17)	539.17***					

Mean of mortality percent ± standard error within a column followed by the same letter did not differ significantly according to Tukey test at P= 0.05; ^{ns}P>0.05; *P<0.05; ***: p<0.001; CI= Confidence interval; SE= Standard error; R²=Coefficient of determination; χ² = Chi-square; Number of replices:4

4. DISCUSSION

Seeds of numerous wild or domestic edible fruits are commonly discarded after its pulps locally consumed or industrially processed. Worldwide, fruit seed wastes (mango, avocado, etc.) are produced in hundreds of thousands tons yearly and their use as medicine, animal food supplements or insecticides may reduce the environmental disposal problems [19]. Indeed, several previous studies reported the insecticidal efficacy of diverse fruit seed extracts and oils against insect pests and particularly against mosquito species. Study carried out on seed ethanolic extracts of 21 Brazilian plants revealed only three plant seed extracts including *Myracrodruon urundeuva*, *Piptadenia moniliformis* and *Luetzelburgia auriculata* having significant insecticidal property against immature stages of *Ae. aegypti* [42]. Methanol seed extracts of *Momordica charantia*, *Azadirachta indica*, and *Ricinus communis*, were found toxic against *Anopheles stephensi* larvae with LC₅₀ values of 87.00, 15.25 and 54.95 ppm, respectively recorded [43]. Seed extracts of rough lemon (*Citrus jambhiri*) and lemon (*Citrus limon*) were revealed being toxic against *Aedes albopictus* with lowest LC₅₀ values of 119.99 and 137.25 ppm respectively registered [44]. The ethanol, chloroform and acetone seed extracts of *Datura stramonium* were found highly effective in the control of rice weevil, *Sitophilus oryzae* [45]. The hexane of *Zanthoxylum heitzii* seed extract played a significant insecticidal efficacy against *An. gambiae* s.s [46]. Acetone, chloroform, and methanol leaf extracts of *M. indica* exhibited a moderate larvicidal activity against *Cx. quinquefasciatus* larvae [34]. *M. indica* crude extract caused substantial activity against *Ae. aegypti* larvae [35]. Against other insect pests, *Tagetes erecta*, *Cynodon dactylon* and *Azadirachta indica* seed extracts showed high direct contact toxicity towards red flour beetles *Tribolium castaneum* [47]. From Bangladesh, seed extracts of *Aphanamixis polystachya* exhibited a strong insecticidal and repellent effects as well as moderate feeding deterrent activity against *Tribolium castaneum* [48]. *Lansium domesticum*, *Annona muricata*, *A. squamosa*, and *Sandoricum koetjape* ethanolic seed extracts significantly inhibited the growth of the polyphagous lepidopteran pest *Spodoptera litura* larvae [49]. The crude seed extracts of *Annona squamosa* was found also toxic and possessed antifeedant effect against larvae of cabbage moths, *Plutella xylostella* and *Trichoplusia ni* [50,51].

Those seed extracts might contain diverse bioactive compounds acting singly or in synergic on mosquito eggs larvae, pupae and adults. Most of these discarded fruit seeds might contain diverse phytochemical constituents such as polyphenols, alkaloids, tannins, steroids, flavonoids, etc., having insecticidal proprieties. Like in this present study, *P. Americana* and *D. edulis* seed extracts were rich in alkaloids, flavonoids, tannins, saponins and phenolic compounds and consequently exhibited a high dose-dependent larvicidal efficacy against *Cx. quinquefasciatus*, *An. gambiae* and *Ae. aegypti* larvae. Similarly, Leite et al. [52] detected condensed tannins, flavonoids, triterpenes and alkaloids in methanol seed extract of avocado and these phytochemicals were responsible of high mortality of *Ae. aegypti* larvae with LC₅₀ value of 8.87 mg.mL⁻¹. The phytochemical β -sitosterol present in the petroleum ether extract of *Abutilon indicum* was reported to be the responsible of high larvicidal activity against *An. stephensi*, *Ae. aegypti*, and *Cx. quinquefasciatus* [53]. The phytochemical screening of *Annona squamosa* and *A. muricata* seed extracts revealed that they are rich in flavonoid and alkaloids and consequently induced high mortality ranging from 0.5 to 1% against larvae and from 1 to 5% against adults of *Ae. albopictus* and *Cx. quinquefasciatus* [54]. Dabas et al. [25] reported the richness of avocado seeds in phenolic compounds, playing a role in the toxic effect against insects. The phytochemical screening of ethanol seed extract of *P. americana* conducted by Torres et al. [26] showed the presence of alkaloids, tannins, saponins, unsaturated steroids and triterpenoids, flavonoids, fats and oils and also exhibited high larvicidal toxicity against *Ae. aegypti* larvae with LC₅₀ of 16.48 mg/L and LC₉₀ of 45.77 mg/L. Some plant components such as the lipids and fatty acids detected in the seed of *D. edulis* contain linoleic acid and linolenic acid, unsaturated fatty acids oleic acid having larvicidal properties against mosquito larvae [28,55]. Study conducted by Torres et al. [26] revealed hexane seed extract of *P. americana* as the most toxic with LC₅₀ of 9.82 mg/L and LC₉₀ of 22.19 mg/L compared to ethanol seed extract of the plant fruit with LC₅₀ of 16.48 mg/L and LC₉₀ of 45.77 mg/L on *Ae. aegypti* larvae. Those authors attributed the highest efficacy of that seed extracts to their richness in alkaloids, tannins, saponins, unsaturated steroids and triterpenoids, flavonoids, steroids, terpenoids, essential oils, fats and oils and phenolic compounds previously reported to for their insecticidal properties [56]. In

fact, inhalation, ingestion or cuticle absorption of plant secondary metabolites by mosquito larvae may create basic metabolic, biochemical, physiological and behavioral dysfunctions in the insect systems [57]. According to Rattan et al. [58], botanical secondary metabolites once in contact or ingurgitate by mosquito larvae may penetrate in the insect system to cause a serial physiological dysfunction such as the inhibition of neurotransmitter synthesis, receptor function or pathway enzymes transduction. For the authors, some phytochemical compounds can inhibit GABA-gated chloride channel, acetylcholinesterase and cellular respiration leading to molecular events disruption causing behavior and memory alteration followed by the weakening and the death of the insect.

Coefficient of determination (R^2) values for the 3 plant seed extracts against larvae of mosquito species were ≥ 0.6 , confirming the effectiveness of our 3 plant seed extracts tested. Indeed, according to Faraway [59], regression analysis model using result data from the biological experiments are favorably attributed to the efficacy of the products if $R^2 \geq 0.6$.

5. CONCLUSION

The three plant seed extracts tested in this present study exhibited a high larvicidal property against *An. gambiae*, *Ae. aegypti* and *Cx. quinquefasciatus* larvae. Overall, the seed extract of *P. Americana* was the most effective against the three mosquito species larvae evaluated. Among the three mosquito species investigated, larvae of *An. gambiae* were the most sensible to all plant seed extracts tested. Thus, the three plant seed extracts and especially *P. americana* seed extract should be considered as a candidate of the new botanical mosquito larvicide to control mosquito larvae in their breeding sites around the buildings.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

ACKNOWLEDGEMENTS

Authors are grateful to the Institute of Medical Research and Medicinal Plant Studies where

plant extraction and phytochemical screenings as well as mosquito larvicidal tests were conducted.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Becker N, Petrić D, Zgomba M, Boase C, Madon M, Dahl C, Kaiser A. Mosquitoes and Their Control 2nd ed. Springer Verlag Berlin Heidelberg. 2010;15:577.
2. Wilder-Smith A, Chen LH, Massad E, Wilson ME. Threat of dengue to blood safety in dengue-endemic countries. Emerging Infectious Diseases. 2009;15:8–11.
DOI:<https://doi.org/10.3201/eid1501.071097>
3. WHO. Monitoring and Epidemiological assessment of mass drug administration in the global programme to eliminate lymphatic filariasis: a manual for national elimination programmes. World Health Organization, Geneva; 2011.
4. Simonsen PE, Mwakitalu ME. Urban lymphatic filariasis. Parasitology Research. 2013;112:35-44.
DOI:<https://doi.org/10.1007/s00436-012-3226-x>
5. WHO. Global programme to eliminate lymphatic filariasis: Progress report, 2019. Weekly epidemiological record. World Health Organization. 2020;95(43):509-524.
6. Nana-Djeunga HC, Tchatchueng-Mbouguia JB, Bopda J, Mbickmen-Tchana S, Elong-Kana N, Nnomzo'o E, Akame J, Njiokou F, Kamgno J. Mapping of bancroftian filariasis in Cameroon: prospects for elimination. PLoS Neglected Tropical Diseases. 2015; 9(9): e0004001.
DOI:<https://doi.org/10.1371/journal.pntd.0004001>
7. Kuniholm MH, Wolfe ND, Huang CY, Mpoudi-Ngole E, Tamoufe U, Le Breton M, Burke DS, Gubler DJ. Seroprevalence and distribution of Flaviviridae, Togaviridae, and Bunyaviridae arboviral infections in rural Cameroonian adults. The American Journal of Tropical Medicine and Hygiene. 2006;74(6):1078-1083.
PMID: 16760524
8. Demanou M, Pouillot R, Grandadam M, Boisier P, Kamgang B, Herve JP, Rogier

- C, Rousset D, Paupy C. Evidence of dengue virus transmission and factors associated with the presence of anti-dengue virus antibodies in humans in three major towns in Cameroon. PLoS Neglected Tropical Diseases. 2014;8(7): e2950.
DOI:https://doi.org/10.1371/journal.pntd.002950
9. Yousseu FBS, Nemg FBS, Ngouanet SA, Mekanda FMO, Demanou M. Detection and serotyping of dengue viruses in febrile patients consulting at the New-Bell District Hospital in Douala, Cameroon. PloS one. 2018;13(10):e0204143.
DOI:https://doi.org/10.1371/journal.pone.0204143
 10. Nemg Simo FB, Sado Yousseu FB, Evouna Mbarga A, Bigna JJ, Melong A, Ntoudé A, Kamgang B, Bouyne R, Moundipa Fewou P, Demanou M: Investigation of an Outbreak of Dengue Virus Serotype 1 in a Rural Area of Kribi, South Cameroon: A Cross-Sectional Study. Intervirology. 2018;61(6):265-271.
DOI:https://doi.org/10.1159/000499128
 11. Monamele GC, Demanou M. First documented evidence of dengue and malaria co-infection in children attending two health centers in Yaounde, Cameroon. The Pan African medical journal. 2018;29: 227.
DOI:https://doi.org/10.11604/pamj.2018.29
 12. Kamgang B, Vazeille M, Yougang AP, Tedjou AN, Wilson-Bahun TA, Mousson L, Wondji CS, Failloux AB. Potential of *Aedes albopictus* and *Aedes aegypti* (Diptera: Culicidae) to transmit yellow fever virus in urban areas in Central Africa. Emerging Microbes & Infections. 2019;8(1):1636-1641.
DOI:https://doi.org/10.1080/22221751.2019.1688097
 13. WHO. World malaria report. World Health Organization, Geneva. Licence: CC BY-NC-SA 3.0 IGO; 2019.
 14. Forattini OP. Espécie de *Culex* (*Culex*). In: Forattini OP, editor. Culicidologia Médica. São Paulo: Editora Universidade de São Paulo. 2002;693–722.
 15. Afzal S, Shah SS, Ghaffar S, Azam S, Arif F. Review on activity of medicinal plant extracts against mosquito genera *Anopheles* & *Culex*. International Journal of Entomology Research. 2018;36:8-14.
 16. Lopes RP, Lima JBP, Martins AJ. Insecticide resistance in *Culex quinquefasciatus* Say, 1823 in Brazil: A review. Parasites & Vectors. 2019;12:591-603.
DOI:https://doi.org/10.1186/s13071-019-3850-8
 17. Miresmailli S, Bradbury R, Isman MB. Comparative toxicity of *Rosmarinus officinalis* L. essential oil and blends of its major constituents against *Tetranychus urticae* Koch (Acari: Tetranychidae) on two different host plants, Pest Management Science. 2006;62:366-371.
DOI:https://doi.org/10.1002/ps.1157
 18. Gutierrez PMJ, Antepuesto AN, Eugenio BAL, Santos MF. Larvicidal activity of selected plant extracts against the dengue vector *Aedes aegypti* mosquito. International Research Journal of Biological Science. 2014;3(4):23-32.
 19. Jedele S, Hau AM, von Oppen M. “An analysis of the world market for mangos and its importance for developing countries”, Deutscher Tropentag, Göttingen. Conference on International Agricultural Research for Development; 2003.
 20. Dabas D, Elias RJ, Lambert JD, Ziegler GR. A colored avocado seed extract as a potential natural colorant. Journal of Food Science. 2011;76(9):C1335–C1341.
DOI:https://doi.org/10.1111/j.1750-3841.2011.02415.x
 21. Hatzakis E, Mazzola EP, Shegog RM, Ziegler GR, Lambert JD. Perseorangin: A natural pigment from avocado (*Persea americana*) seed. Food Chemistry. 2019; 293:15-22.
DOI:https://doi.org/10.1016/j.foodchem.2019.04.064
 22. Adeyemi OO, Okpo SO, Ogunti OO. Analgesic and anti-inflammatory effects of the aqueous extract of leaves of *Persea americana* Mill (Lauraceae). Fitoterapia. 2002;73:375-380.
DOI:https://doi.org/10.1016/s0367-326x(02)00118-1
 23. Ozolua RI, Anaka ON, Okpo SO, Idogun SE. Acute and sub-acute toxicological assessment of the aqueous seed extract of *Persea americana* Mill (Lauraceae) in rats. African Journal of Traditional Complementary and Alternative Medicine. 2009; 6(4):573–578.
DOI:https://doi.org/10.4314/ajtcam.v6i4.57214

24. Lima MLR, Pushpa V, Balakrishna K, Ganesan P. Mosquito larvicidal activity of Avocado (*Persea americana* Mill.) unripe fruit peel methanolic extract against *Aedes aegypti*, *Culex quinquefasciatus* and *Anopheles stephensi*. South African Journal of Botany. 2020;133:1-4. DOI:<https://doi.org/10.1016/j.sajb.2020.06.020>
25. Dabas D, Shegog RM, Ziegler GR, Lambert JD. Avocado (*Persea americana*) seed as a source of bioactive phytochemicals. Current Pharmaceutical Design. 2013;19(34):6133-6140. DOI:<https://doi.org/10.2174/1381612811319340007>
26. Torres RC, Garbo AG, Walde RZM. Larvicidal activity of *Persea americana* Mill. against *Aedes aegypti*. Asian Pacific Journal of Tropical Medicine. 2014;7(1): S167-S170. DOI:[https://doi.org/10.1016/S1995-7645\(14\)60225-X](https://doi.org/10.1016/S1995-7645(14)60225-X)
27. Omogbai BA, Ojeaburu SI. Nutritional composition and microbial spoilage of *Dacryodes edulis* fruits vended in Southern Nigeria. Science World Journal. 2010;5(4): 5-10. Available: www.scienceworldjournal.org ISSN 1597-6343
28. Ajibesin KK. *Dacryodes edulis* (G. Don) H. J. Lam: A review of its medicinal, phytochemical and economic properties. Research Journal of Medicinal Plants. 2011; 5: 32-41. DOI:<https://doi.org/10.3923/rjmp.2011.32.41>
29. Omonhinmin AC, Agbara UI. Assessment of In vivo antioxidant properties of *Dacryodes edulis* and *Ficus exasperate* as anti-malaria plants. Asian Pacific Journal of Tropical Disease. 2011;3(4):294-300. DOI:[https://doi.org/10.1016/S2222-1808\(13\)60072-9](https://doi.org/10.1016/S2222-1808(13)60072-9)
30. Omogbai BA, Eneh TO. Antibacterial activity of *Dacryodes edulis* seed extract on food borne diseases. Bayero Journal of Pure and Applied Sciences. 2011;4(1):17-21. DOI:<https://doi.org/10.4314/bajopas.v4i1.3>
31. Oladimeji OH, Nia R, Nyong E, Kalu N. Evaluation of larvicidal and antimicrobial potential of *Dacryodes edulis* G. Don; H.J. Lam (Burseraceae). Journal of Pharmacy & Bioresources. 2010;7(2). DOI:<https://doi.org/10.4314/jpb.v7i2.8>
32. Akande-Grillo HT, Nzelibe HC. Larvicidal activity of extracts of *Dacryodes edulis* (G. Don) H. J Lam on *Aedes vittatus* Bigot and *Culex quinquefasciatus* Say (Diptera: Culicidae). International Journal of Mosquito Research. 2017;4(4):32-36. ISSN: 2348-5906, CODEN: IJMRR2
33. Orwa C, Mutua A, Kindt R, Jamnadass R, Anthony S. Agroforestry Database: A tree reference and selection guide version 4.0", World Agroforestry Centre, Kenya; 2009.
34. Rahuman AA, Bagavan A, Kamaraj C, Vadivelu M, Abdus Zahir A, Elango G, Pandiyan G. Evaluation of indigenous plant extracts against larvae of *Culex quinquefasciatus* Say (Diptera: Culicidae). Parasitology Research. 2009;104(3):637-643. DOI:<https://doi.org/10.1007/s00436-008-1240-9>
35. Mohammed A, Chadee DD. An evaluation of some Trinidadian plant extracts against larvae of *Aedes aegypti* mosquitoes," Journal of the American Mosquito Control Association. 2007;23(2):172-176. DOI:[https://doi.org/10.2987/8756-971X\(2007\)](https://doi.org/10.2987/8756-971X(2007))
36. Harborne JB. Textbook of phytochemical Methods. Guide to modern techniques of plant analysis 5 th Edition, Chapman and Hall Ltd, London. 2008;21-72.
37. Azari-Hamidian S, Harbach RE. Keys to the adult females and fourth instar larvae of mosquitoes of Iran (Diptera: Culicidae). Zootaxa. 2009;2078:1-33. DOI:<https://doi.org/10.5281/zenodo.187282>
38. Gillies MT, Coetzee M. A supplement to the Anophelinae of Africa south of the Sahara. South African Institute of Medical Research. 1987;55:143.
39. WHO. Guidelines for Laboratory and Field Testing of Mosquito Larvicides, World Health Organization, Geneva, Switzerland; 2005.
40. Abbott WS. A method of computing the effectiveness of an insecticide. Journal of Economic Entomology. 1925;18:265-266.
41. Finney DJ. Probit analysis. 3rd edition, Cambridge University Press, London. 1971;68-72.
42. Souza TM, Farias DF, Soares BM, Viana MP, Lima GPG, Machado LKA, Morais SM, Carvalho AFU. Toxicity of brazilian plant seed extracts to two strains of *Aedes aegypti* (Diptera: Culicidae) and Non-target

- Animals. Journal of Medical Entomology. 2011;48(4):846–851.
DOI:<https://doi.org/10.1603/me10205>
43. Batabyal L, Sharma P, Mohan L, Maurya P, Srivastava CN. Larvicidal efficiency of certain seed extracts against *Anopheles Stephensi*, with reference to *Azadirachta indica*. Journal of Asia-Pacific Entomology. 2007;10(3):251-255.
DOI:[https://doi.org/10.1016/S1226-8615\(08\)60359-3](https://doi.org/10.1016/S1226-8615(08)60359-3)
 44. Akram W, Ali Khan HA, Hafeez F, Bilal H, Kim YK, Lee JJ. Potential of Citrus seed extracts against dengue fever mosquito, *Aedes albopictus* (Skuse) (Culicidae: Diptera). Pakistan Journal of Botany. 2010; 42(4):3343-3348.
ISBN0620032x3
 45. Jawalkar N, Zambare S, Zanke S. Insecticidal property of *Datura stramonium* L. seed extracts against *Sitophilus oryzae* L. (Coleoptera: Curculionidae) in stored wheat grains. Journal of Entomology and Zoology Studies. 2006;4(6):92-96.
Corpus ID: 55167783
 46. Overgaard HJ, Sirisopa P, Mikolo B, Malterud KE, Wangensteen H, Zou Y-F, Paulsen BS, Massamba D, Duchon S, Corbel V, Chandre F. Insecticidal Activities of Bark, Leaf and Seed Extracts of *Zanthoxylum heitzii* against the African Malaria Vector *Anopheles gambiae*. *Molecules*. 2014;19:21276-21290.
DOI:<https://doi.org/10.3390/molecules191221276>
 47. Islam MS, Talukder FA. Toxic and residual effects of *Azadirachta indica*, *Tagetes erecta* and *Cynodon dactylon* seed extracts and leaf powders towards *Tribolium castaneum*. Journal of Plant Diseases and Protection. 2005;112(6): 594–601.
DOI:<https://doi.org/10.1007/BF03356157>
 48. Talukder FA, Howse PE. Deterrent and insecticidal effects of extracts of pithraj, *Aphanamixis polystachya* (Meliaceae), against *Tribolium castaneum* in storage. Journal of Chemical Ecology. 1993;19: 2463–2471.
DOI:<https://doi.org/10.1007/BF00980683>
 49. Leatemia JA, Isman MB. Insecticidal activity of crude seed extracts of *Annona* spp., *Lansium domesticum* and *Sandoricum koetjape* against Lepidopteran larvae. *Phytoparasitica*. 2004;32(1):30-37.
DOI:<https://doi.org/10.1007/BF02980856>
 50. Leatemia JA, Isman MB. Toxicity and antifeedant activity of crude seed extracts of *Annona squamosa* (Annonaceae) against lepidopteran pests and natural enemies. International Journal of Tropical Insect Science. 2004;24:150–158.
DOI:<https://doi.org/10.1079/IJT200416>
 51. Leatemia JA, Isman MB. Efficacy of crude seed extracts of *Annona squamosa* against diamondback moth, *Plutella xylostella* L. in the greenhouse. International Journal of Pest Management. 2004;50(2):129-133.
DOI :<https://doi.org/10.1080/09670870410001691821>
 52. Leite JJG, Brito EHS, Cordeiro RA, Brilhante RSN, Sidrim JJC, Bertini LM, de Morais SM, Rocha MFG. Chemical composition, toxicity and larvicidal and antifungal activities of *Persea americana* (avocado) seed extracts. *Revista da Sociedade Brasileira de Medicina Tropical*. 2009; 42(2):110-113.
DOI:<http://dx.doi.org/10.1590/S0037-86822009000200003>
 53. Rahuman AA, Gopalakrishnan G, Venkatesan P, Geetha K. Isolation and identification of mosquito larvicidal compound from *Abutilon indicum* (Linn) Sweet. *Parasitology Research*. 2008;102: 981-988.
DOI:<https://doi.org/10.1007/s00436-007-0864-5>
 54. Ravaomanarivo LHR, Razafindraleva HA, Raharimalala FN, Rasoahantaveloniaina B, Ravelonandro PH, Mavingui P. Efficacy of seed extracts of *Annona squamosa* and *Annona muricata* (Annonaceae) for the control of *Aedes albopictus* and *Culex quinquefasciatus* (Culicidae). *Asian Pacific Journal of Tropical Biomedicine*. 2014; 4(10):798-806.
DOI:<https://doi.org/10.12980/APJTb.4.2014C1264>
 55. Haribalan P, Yung JJ, Jun-Ran K, Murugan K, Young-Joon A. Larvicidal activity and possible mode of action of flavonoids and two fatty acids identified in *Millettia pinnata* seed towards three mosquito species. *Parasites and Vectors*. 2015;8:237.
DOI:<https://doi.org/10.1186/s13071-015-0848-8>
 56. Shaalan EA, Canyon D, Younes MW, Abdel-Wahab H, Mansour AH. A review of

- botanical phytochemicals with
mosquitocidal potential. Environment
International. 2005;31(8):1149-1166.
DOI:<https://doi.org/10.1016/j.envint.2005.03.003>
57. Khater HF. Ecosmart Biorational
Insecticides: Alternative insect control
strategies, insecticides. Advances in
Integrated Pest Management, In Tech.
2012:708.
DOI:<https://doi.org/10.5772/27852>
58. Rattan RS. Mechanism of action of
insecticidal secondary metabolites of plant
origin. Crop Protection. 2010;29:913-920.
59. Faraway JJ. Practical Regression and
Anova using R. 2002;212.

© 2020 Younoussa et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
<http://www.sdiarticle4.com/review-history/64536>