



A Review on Antiurolithiatic (Pashanabeda) Effects of Herbal and Marine Resources

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Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JPRI/2021/v33i63A36096

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/85410>

Received 20 October 2021

Accepted 28 December 2021

Published 29 December 2021

Review Article

ABSTRACT

Urolithiasis (urinary calculi) is stone development in bladder, urinary tract and kidney. The rise in frequency and occurrence of kidney stones is talking point all over the world. 12% of the global population suffer from urinary stone formation. The phenomenon effect of urolithiasis is lowered by using the pashanabeda elements. Pashanabeda (Pashan: stone; bheda: to break) is a Sanskrit locution which implies the breakage of stone. The adverse reactions caused due to man made drugs has prompted to rejoin with the natural safe medication. The evolution of plant-based therapeutics has shown terrific notice and demand as an modernistic drug entities all over the sphere. The present review aims the investigators to easily identify and develop plant and marine resources beneficial in management of urolithiasis.

Keywords: Urolithiasis; antiurolithiatic; pashanabeda; herbal; animal; marine; microbial; mineral.

1. INTRODUCTION

Humankind is known to be suffering from urinary stone disease since ancient times, and it was first found in tombs of Egyptian mummies dating back to 4000BC and in the graves of North American Indians from 1500 BC – 1000BC [1].

2. UROLITHIASIS OR NEPHROLITHIASIS

Urolithiasis or nephrolithiasis is one of the oldest and endemic painful urological disorder [2-10]. The genesis of stone in urinary system i.e., in the kidney, ureter and urinary bladder or in the

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urethra is known as urolithiasis (ouron =urine, lithos = stone) [11].

World population of 3-20% have propensity to form single urinary stone in their lifespan of 70 years [12]. 12% of global population suffer from urinary stone formation [13].

The average lifetime risk of stone formation has been reported in the range of 5-10% population in which there is a predominance of men over women. A handful of lithogenic factors that leads to formation of stones such as sexuality, age,

profession, food habits, water intake, education level, national diffusion, socioeconomic status, metabolic and genetic disorders and diseases (Mohammad Shazib Faridi et al. 2020); [14,15].

3. MECHANISM OF URINARY STONE FORMATION

Stone genesis is complicated process which results because of succeeding Physicochemical occurrences such as excellent saturation, growth, nucleation, retention and aggregation within the renal tubules.

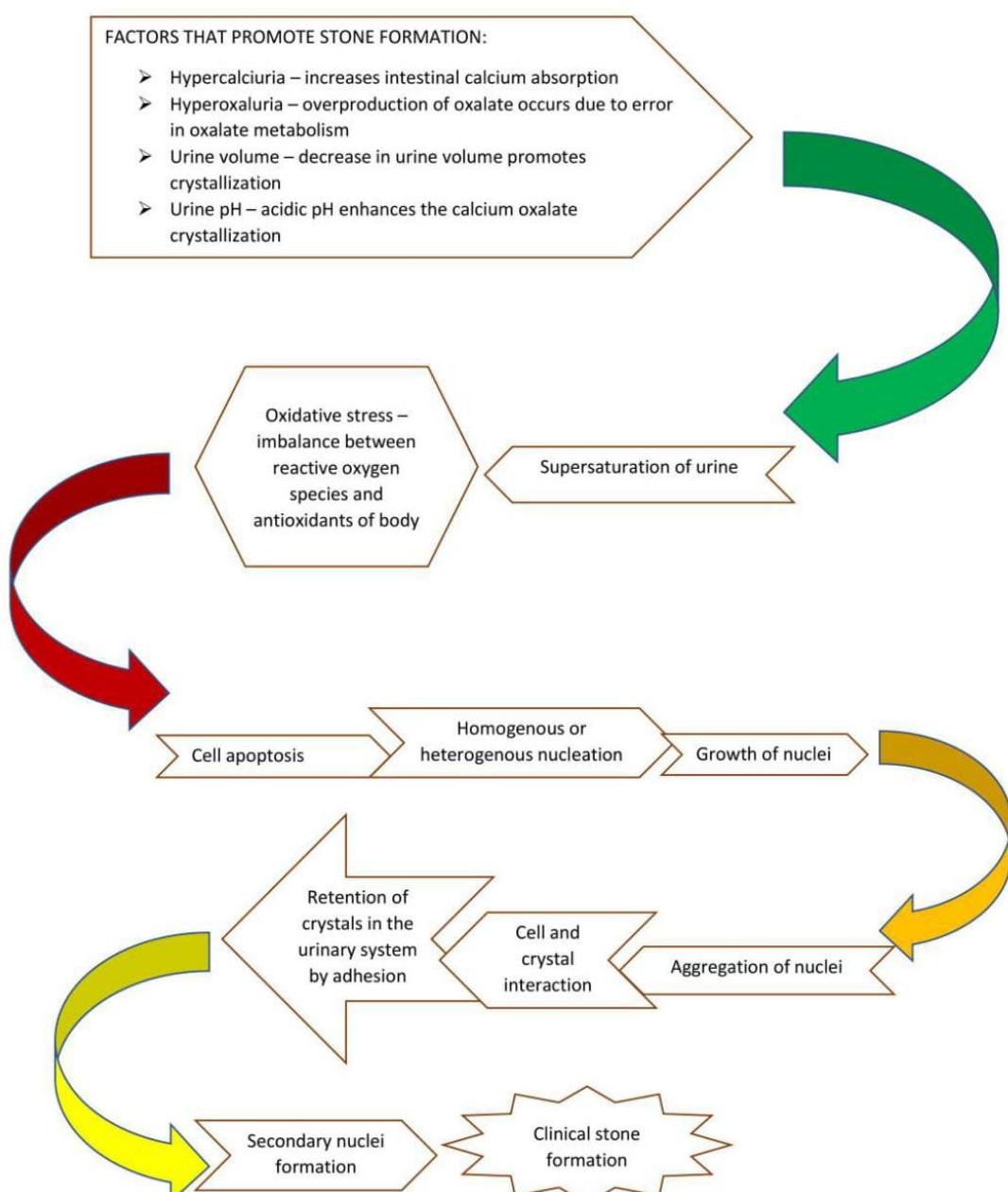


Fig. 1. Flowchart of mechanism of urinary stone formation

The mechanism of stone formation comprises seedling of stone crystals (nuclei formation, nucleation), assemblage of nuclei to interact with inter renal structure (aggregation) and thus maintain themselves in renal system further aggregate to form secondary nuclei which shape into clinical stone (crystal development) [16].

Constituents of stone components may differ, but the severity it causes may vary based on its site, severity of stone formation and its action at site.

The mechanism of stone formation is depicted in flowchart format as indicated below.

3.1 Types of Urolithiasis

The kidney stone size varies from 5mm to 7mm. different types of kidney stones are included in table format. The type of stone formed is named after its mineral composition. The most frequent stones are struvite (magnesium ammonium

phosphate), calcium oxalate, urate, cysteine and silica [13].

3.2 Pashanabhedha

Pashanabeda is a Sanskrit phrase where the word Pashan means stone and bheda means to break, which literally means the breakage of stones. Several plants such as Bergenia ligulata syn., Saxifraga ligulata etc. are denominated as pashanabeda as they show properties similar to that of water pills and lithiotriptic activities which is included in classics of Ayurveda. (www.ccras.nic.in) [17].

3.3 Antiurolithiatic Activity

The components that reduce the stone shaping components in urine and reduce renal retention are known as antiurolithiatic components. The activity is known as antiurolithiatic activity.

Table 1. Types of kidney stones [13]

S. No.	Types of kidney stones	Percentage of occurrence	Causes for kidney stones	Characteristic feature of stones
1.	Calcium stones	-	Calcium combines with oxalate and phosphate to form calcium oxalate and calcium phosphate stones. The rise in oxalate and phosphate levels in dietary supplements leads to formation of stones	White, black or grey colored components, radio opaque
2.	Struvite stones	10-15%	Stones are formed due to bacterial infections	Large, glared and laminated
3.	Uric acid stones	5-10%	People with malformation like gout syndrome, obesity and food source rich in purine, protein (meat & fish) results in stone formation	Yellow orange colored, smooth, round or square or diamond or rod shape, pleomorphic
4.	Protease related stones	4-12%	Formation of crystals are recognized in HIV positive patients using protease inhibitors (indinavir sulphate)	-
5.	Cysteine stones	-	Hereditary disorders induce formation of stones	Greenish yellow color, round, shiny, radiopaque
6.	Silica stones	-	Synthetic and herbal medicines induce formation of stones. These are crystal called as drug induced stones	-

This review article mainly focuses on various plants and marine accredited with diuretic and antiurolithiatic activities.

4. METHODOLOGY

The components are classified into natural sources and synthetic sources. Natural sources are further classified into herbal source, animal source, marine source, microbial source and mineral source.

4.1 Herbal Sources

The plants that show antiurolithiatic activity and diuretic activity are classified based on their

morphological characteristics and depicted examples in alphabetical order. It is further depicted in the form of table as shown below (Table 2) [18].

4.2 Animal Source

The animal source with litholytic activity are listed in Table 3 [1].

4.3 Marine Sources

The marine source with antiurolithiatic activity and diuretic property are listed in Table 4 [1].

Table 2. Plants with antiurolithiatic and diuretic activity

S.No.	Class (Root)	Antiurolithiatic plants
1.	<i>Abutilon muticum</i> (Delile ex DC.) <i>Achyranthus aspera</i> L. <i>Acorus calamus</i> <i>Aerva javanica</i> <i>Ageratum conyzoides</i>	<i>Hygrophila auriculata</i> <i>Hygrophila schulli</i> <i>Ichnocarpus frutescens</i> (L.) W.T.Aiton <i>Indigofera tinctoria</i> <i>Juniperus chinensis</i> . <i>Juniperus polycarpos</i> K.Koch
	<i>Alcea fasciculiflora</i> Zohary <i>Alhagi manniifera</i> Jaub. & Spach <i>Alhagi maurorum</i> Medik. <i>Alisma orientale</i> (Sam.) Juz. <i>Angelica sinesis polysaccharide</i> <i>Apium graveolens</i> L. <i>Aquilegia fragrans</i> Benth. <i>Arctium lappa</i> <i>Arnebia euchroma</i> <i>Asclepias syriaca</i> L. <i>Asparagus officinalis</i> <i>Asparagus racemosus</i> <i>Asparagopsis adscendens</i> Kunth. / <i>Asparagus officinalis</i> <i>Anneslea fragrans</i> Wall. / <i>Aquilegia fragrans</i> Benth. <i>Barbaarea vulgaris</i> R.Br. <i>Barleria prionitis</i> L. <i>Berberis vulgaris</i> <i>Bergenia ciliata</i> <i>Beta vulgaris</i> L. <i>Biophytum abyssinicum</i> Steud. Ex A. Rich. <i>Biophytum reinwardtii</i> <i>Biophytum sensitivum</i> L. <i>Boerhavia diffusa</i> <i>Caesalpinia nuga</i> (Aiton.) <i>Celosia argentea</i> <i>Ceterach aureum</i> Buch. <i>Chondrodendron tomentosum</i> Ruiz & Pav. <i>Cissampelos parerira</i> L. <i>Citrus limon</i> (L.) Osbeck <i>Clerodendrum serratum</i>	<i>Lawsonia inermis</i> L. <i>Levisticum officinale</i> <i>Medicago sativa</i> L. <i>Mimosa pudica</i> L. <i>Moringa oleifera root-wood</i> <i>Musa balbisiana</i> Colla. <i>Musa x paradisiaca</i> L. <i>Nothosaerva brachiate</i> (L.) Wight. <i>Ononis spinosa</i> <i>Petroselinum crispum</i> <i>Petroselinum sativum</i> <i>Phragmites australis</i> (Cav.) Trin. Ex Steud. <i>Piper methysticum</i> G.Forst. <i>Plantago major</i> L. <i>Raphanus sativus</i> L. <i>Rheum emodi</i> Wall. <i>Ribes triste</i> Pall. <i>Ricinus communis</i> L. <i>Rotula aquatica</i> Lour. <i>Rubia tinctorum</i> <i>Rubus caesius</i> L. <i>Rubus fruticosus</i> Lour. <i>Rubus sanctus</i> Schreb. <i>Rumex acetosella</i> <i>Rumex hastatus</i> <i>Saccharum officinarum</i> <i>Saccharum spontaneum</i> <i>Sageretia brandrethiana</i> Aitch. <i>Saponaria mesogitana</i> Boiss.

S.No.	Class (Root)	Antiulrolithic plants
	<i>Clitoria ternatea</i> <i>Coccinia grandis</i> <i>Coix lacryma-jobi</i> <i>Costus arabicus</i> <i>Costus igneus</i> <i>Crotalaria albida</i> Roth. <i>Crotalaria pallida</i> Aiton. <i>Cucumis sativus</i> L. <i>Cynara scolymus</i> L. <i>Cynodont dactylon</i> (L.) Pers. <i>Daucus carota</i> L. <i>Ecbolium viride</i> (Forssk.) Alston <i>Echinops spinosus</i> L. <i>Ensete superbum</i> (Roxb.) Cheesman. <i>Eryngium creticum</i> Lam. <i>Eupatorium purpureum</i> L. <i>Filipendula vulgaris</i> Moench <i>Glycyrrhiza glabra</i> L. <i>Gypsophila struthium</i> Loefl. <i>Hamelia patens</i> Jacq. <i>Hemidesmus indicus</i> <i>Helianthus annuus</i> <i>Homonia riparia</i> Lour. <i>Holarrhena antidysenterica</i> <i>Hydrangea arborescens</i> L.	<i>Scoparia dulcis</i> L. <i>Sida rhombifolia</i> <i>Smilax aspera</i> <i>Solanum anguivi</i> Lam. <i>Solanum incanum</i> L. <i>Solanum surattense</i> Burm.f. <i>Solanum virginianum</i> L. <i>Sphaeranthus indicus</i> L. <i>Strychnos potatorum</i> L.f. <i>Taraxacum officinale</i> <i>Tectona grandis</i> <i>Tephrosia purpurea</i> <i>Tinospora purpurea</i> <i>Traxacum pseudobrachyglossum</i> Soest. <i>Trianthema portulacastrum</i> <i>Tribulus terrestris</i> <i>Tropaeolum tuberosum</i> Ruiz & Pav <i>Valeriana officinalis</i> L. <i>Valeriana wallichii</i> DC. <i>Vetiveria zizanioides</i> <i>Vitex negundo</i> <i>Xanthium strumarium</i> <i>Xenostegia tridentata</i> Webb & Berthel <i>Zaleya pentandra</i> (L.) C.Jeffrey <i>Ziziphus lotus</i> (L.) Lam.
2.	Rhizome <i>Agropyron repens</i> (L.) P. Beauv. <i>Asparagus racemosus</i> Willd <i>Bergenia ligulata</i> Engl. <i>Beta vulgaris</i> L. <i>Colocasia esculenta</i> (L.) Schott. <i>Curculigo orchoides</i> Gaertn.	<i>Cynodon dactylon</i> <i>Elymus repens</i> <i>Eupatorium purpureum</i> <i>Hedychium coronarium</i> J. Koenig. <i>Smilax lanceifolia</i> Roxb. <i>Zingiber officinale</i> Roscoe
3	Bulb <i>Allium cepa</i> L. <i>Allium sativum</i> L.	<i>Drimia indica</i> (Roxb.) Jessop <i>Scilla indica</i> Roxb.
4	Aerial Parts <i>Achillea falcata</i> L. <i>Arnica montana</i> L. <i>Cardamine uliginosa</i> M.Bieb. <i>Chimaphila maculata</i> (L.) Pursh. <i>Chimaphila umbellata</i> (L.) Nutt. <i>Coccus hirsutus</i> (L.) W.Theob. <i>Crataegus monogyna</i> <i>Crataegus pentagyna</i> <i>Cyperus longus</i> <i>Derris trifoliata</i> Lour. <i>Equisetum ramosissimum</i> <i>Equisetum telmateia</i> .	<i>Gymnocarpus decandrum</i> Forssk. <i>Geranium robertianum</i> L. <i>Helichrysum plicatum</i> DC. <i>Herniaria glabra</i> L. <i>Hyparrhenia hirta</i> (L.) Stapf. <i>Lolium perenne</i> L. <i>Melilotus officinalis</i> (L.) Pall. <i>Orthosiphon aristatus</i> (Blume) Miq. <i>Prosopis farcta</i> (Banks & Sol.)J.F.Macbr. <i>Salvia canariensis</i> <i>Verbena officinalis</i> L. <i>Zilla spinosa</i> (L.) Prantl.
5	Bark <i>Ammi visnaga</i> (L.) Lam. <i>Bauhinia purpurea</i> L. <i>Betula pendula</i> Roth. <i>Berberis vulgaris</i> L. <i>Cinnamomum aromaticum</i> Nees. <i>Cinnamomum bejolghota</i> (Buch. – Ham.)	<i>Juniperus communis</i> <i>Laurus nobilis</i> L. <i>Lawsonia inermis</i> L. <i>Macaranga peltate</i> <i>Mimusops elengi</i> L. <i>Moringa pterygosperma</i> Gaertn

S.No.	Class (Root)	Antiulrolithiatic plants
	Sweet	
	<i>Cinnamomum verum</i> J. Presl.	<i>Neolamarckia cadamba</i> (Roxb.) Bosser.
	<i>Clitoria ternatea</i> L.	<i>Olea europaea</i> L.
	<i>Cedrus deodara</i> (Roxb. Ex D. Don) G.	<i>Picea mariana</i> (Mill.) Britton, Sterns & Poggenb
	<i>Crateva adansonii</i> subsp	<i>Raphanus sativus</i> L.
	<i>Crataeva magna</i> (Lour.) DC	<i>Saraca asoca</i> (Roxb.) Willd.
	<i>Crataeva nurvala</i>	<i>Sonchus oleraceus</i> (L.) L.
	<i>Holarrhena pubescens</i>	<i>Terminalia arjuna</i>
	<i>Hydrangea arborescens</i>	<i>Ziziphus lotus</i>
6	Stem	
	<i>Achyranthus aspera</i> L.	<i>Hedychium aurantiacum</i> Roscoe.
	<i>Butea monosperma</i> (Lam.) Taub.	<i>Musa x paradisiaca</i>
	<i>Bryonia alba</i> L.	<i>Silybum marianum</i> (L.) Gaertn.
	<i>Eryngium campsetre</i> L.	<i>Ruscus aculeatus</i> L.
	<i>Equisetum arvense</i> L.	<i>Tinospora cordifolia</i> (Willd.) Miers.
7	Bamboo Shoots	<i>Bambusa nutans</i> Wall.ex Munro
8	Leaves	
	<i>Aaronsohnia pubescens</i> (Desf.) K.Bremer & Humphries.	<i>Hypericum hypericoides</i>
	<i>Adiantum capillus-veneris</i>	<i>Ilex aquifolium</i> L.
	<i>Allium odorum</i> L.	<i>Larrea tridentata</i>
	<i>Alternanthera brasiliiana</i>	<i>Launaea procumbens</i> L.
	<i>Althaea officinalis</i> L.	<i>Lavandula stoechas</i> L.
	<i>Amaranthus blitum</i>	<i>Lepidium latifolium</i> L.
	<i>Amaranthus caudatus</i>	<i>Lithospermum officinale</i> L
	<i>Ammi visnaga</i> (L.) Lam.	<i>Melia azadirachta</i> L.
	<i>Anisotes trisulcus</i> (Forssk.) Nees	<i>Mentha arvensis</i> .
	<i>Anacardium occidentale</i> L.	<i>Mentha spicata</i> .
	<i>Anneslea fragrans</i> Wall. / <i>Aquilegia fragrans</i> Benth.	<i>Mimosa pudica</i> L.
	<i>Arctostaphylos pungens</i>	<i>Moringa oleifera</i> Lam.
	<i>Arctosyaphylos uva ursi</i>	<i>Musa x paradisiaca</i> L.
	<i>Argyreia nervosa</i> (Burm. f.) Bojer.	<i>Ocimum sanctum</i>
	<i>Asphodelus tenuifolius</i> Cav	<i>Ocimum tenuiflorum</i> L.
	<i>Asplenium ceterach</i> L.	<i>Oenothera biennis</i> L.
	<i>Asplenium hemionitis</i> L.	<i>Olea europaea</i>
	<i>Asplenium scolopendrium</i> L.	<i>Oxalis corniculate</i> L.
	<i>Arum rupicola</i> Boiss.	<i>Paronychia argentea</i> Lam.
	<i>Azadirachta indica</i> A. Juss.	<i>Paederia foetida</i> L.
	<i>Basella alba</i> L.	<i>Peperomia pellucida</i> (L.) Kunth
	<i>Barbara vulgaris</i> R.Br.	<i>Phyllanthus niruri</i>
	<i>Bauhinia forficata</i> Link	<i>Phyllanthus lanceolatus</i> Poir.
	<i>Berberis integerrima</i> Bunge.	<i>Picea smithiana</i> (Wall.) Boiss.
	<i>Betula pendula</i> Roth.	<i>Piper aduncum</i> L.
	<i>Betula lenta</i> L	<i>Piper longum</i> L
	<i>Betula utilis</i> D Don.	<i>Plantago lanceolata</i> L.
	<i>Biophytum abyssinicum</i> Steud. Ex A. Rich.	<i>Plectranthus amboinicus</i> (Lour.) Spreng.
	<i>Boldoa purpurascens</i> cav	<i>Polygonum cognatum</i> Meisn.
	<i>Brassica napus</i> L.	<i>Populus alba</i> L.
	<i>Bryophyllum calycinum</i> Salisb.	<i>Portulaca oleracea</i>
	<i>Cassia auriculata</i> L.	<i>Prosopis farcta</i>
	<i>Calendula officinalis</i> L.	<i>Pulmonaria officinalis</i> L.
	<i>Carissa opaca</i> Stapf ex Haines.	<i>Raphanus sativus</i> L
	<i>Celastrus paniculatus</i>	<i>Rosa canina</i> L.
	<i>Celtis timorensis</i> .	<i>Rosmarinus</i>

S.No.	Class (Root)	Antiulrolithic plants
	<i>Chenopodium album</i> L. <i>Chimaphila maculata</i> (L.) Pursh. <i>Cichorium intybus</i> L. <i>Cissus adnata</i> Roxb. <i>Cissus gongylodes</i> (Baker) <i>Coleus amboinicus</i> Lour <i>Corallodiscus lanuginosus</i> <i>Coriandrum sativum</i> L. <i>Crataegus aronia</i> (L.) Bosc ex DC. <i>Cucumis sativus</i> L. <i>Cyclea peltata</i> (Lam.) Hook.f. & Thomson <i>Didymocarpus pedicellatus</i> R.Br. <i>Duranta erecta</i> <i>Ecbolium viride</i> (Forssk.) Alston <i>Eleusine indica</i> Linn. (poaceae) <i>Enicostema axillare</i> (Poir. ex Lam.) A. Raynal. <i>Eriobotrya japonica</i> (Thunb.) Lindl. <i>Eupatorium birmanicum</i> DC. <i>Ficus carica</i> L. <i>Fraxinus excelsior</i> L. <i>Forsskaolea tenacissima</i> L. <i>Glechoma hederaceae</i> L. <i>Hedera helix</i> L.	<i>Rubus caesius</i> <i>Ruscus aculeatus</i> L. <i>Ruta graveolens</i> L. <i>Sapium sebiferum</i> (L.) Roxb. <i>Saponaria mesogitana</i> Boiss. <i>Smilax aspera</i> L. <i>Solidago virgaurea</i> L. <i>Spergularia rubra</i> <i>Spermacoce hispida</i> L <i>Stachytarpheta indica</i> (L.) Vahl. <i>Tephrosia purpurea</i> (L.) Pers. <i>Teucrium polium</i> L <i>Thunbergia alata</i> Bojer ex Sims. <i>Thymus kotschyanus</i> <i>Thymus migricus</i> <i>Tinospora purpurea</i> (L.) Pers. <i>Tinospora cordifolia</i> (Willd.) Miers. <i>Tournefortia acuminata</i> A.DC. <i>Trapogon buphtalmoides</i> (DC.) Boiss. <i>Trianthema portulacastrum</i> L <i>Tribulus terrestris</i> L. <i>Triclisia gilletti</i> <i>Vitis vinifera</i>
9	Flowers <i>Alcea pallida</i> (Willd.) Waldst. & Kit. <i>Anthemis nobilis</i> L. <i>Borassus flabellifer</i> L. <i>Cassia occidentalis</i> L. <i>Calendula officinalis</i> L. <i>Cocos nucifera</i> L. <i>Cymbopogon schoenanthus</i> (L.) Spreng. <i>Echinops spinosus</i> L. <i>Eryngium campestre</i> L. <i>Helichrysum arenarium</i> (L.) Moench <i>Helichrysum plicatum</i> DC. Subsp. <i>plicatum</i> <i>Hibiscus sabdariffa</i> L.	<i>Inula oculus-christi</i> L <i>Matricaria chamomilla</i> L. <i>Moringa oleifera</i> Lam. <i>Musa x paradisiaca</i> L. <i>Opuntia ficus-indica</i> (L.) Mill. <i>Panicum miliaceum</i> L. <i>Phlogacanthus thyrsiformis</i> Hardow (Mabb) <i>Rosa canina</i> L. (rose hips) <i>Rosa indica</i> L. <i>Rubus fruticosus</i> Lour. <i>Solidago virgaurea</i> L. <i>Taraxacum hybernum</i> Steven <i>Zea mays</i> L. (tea of corn silk)
10	Fruits <i>Ammi visnaga</i> (L.) Lam. <i>Aegle marmelos</i> (L.) Correa <i>Ananas comosus</i> (L.) Merr. <i>Artemisia abrotanum</i> L. <i>Averrhoa carambola</i> L. <i>Ananas comosus</i> (L.) Meerr. <i>Benincasa hispida</i> (Thunb.) Cogn. <i>Bombax ceiba</i> L. <i>Brassica oleracea</i> L. <i>Bunium persicum</i> <i>Cannabis sativa</i> L <i>Cassia fistula</i> L. <i>Cordia ecalyculata</i> Vell. <i>Cordia grandis</i> Roxb. <i>Citrullus vulgaris</i> <i>Cucumis melo</i>	<i>Neolamarckia cadamba</i> <i>Nigella sativa</i> <i>Paeonia officinalis</i> <i>Peganum harmala</i> <i>Pedalium murex</i> <i>Peucedanum grande</i> C.B.Clarke. <i>Phillyrea latifolia</i> L. <i>Phyllanthus emblica</i> L. <i>Physalis alkekengi</i> L <i>Pinus brutia</i> Ten <i>Pinus eldarica</i> Medw. <i>Piper cubeba</i> <i>Piper longum</i> . <i>Piper nigrum</i> <i>Pimpinella anisum</i> <i>Platanus orientalis</i>

S.No.	Class (Root)	Antiulrolithiatic plants
	<i>Cucumis sativus</i>	<i>Prunus avium</i>
	<i>Elettaria cardamomum (L.) Maton.</i>	<i>Punica granatum</i>
	<i>Emblica officinalis Gaertn</i>	<i>Rosa canina</i>
	<i>Gmelina arborea Roxb.</i>	<i>Rhus succedanea L.</i>
	<i>Gossypium herbaceum L.</i>	<i>Ruscus hypoglossum L.</i>
	<i>Ficus carica L.</i>	<i>Rubus ellipticus</i>
	<i>Ficus palmata Forssk.</i>	<i>Rubus fruticosus</i>
	<i>Foeniculum vulgare Mill.</i>	<i>Serenoa repens (W.Bartram) Small</i>
	<i>Juniperus pseudosabina Fisch. & C.A.Mey.</i>	<i>Solanum xanthocarpum</i>
	<i>Kigelia pinnata (Jacq.) DC.</i>	<i>Spondias axillaris Roxb.</i>
	<i>Lagenaria siceraria (Molina) Standl.</i>	<i>Syzygium cumini</i>
	<i>Levisticum officinale</i>	<i>Tamarindus indica L.</i>
	<i>Lithospermum officinale</i>	<i>Tribulus terrestris L.</i>
	<i>Manilkara zapota</i>	<i>Trigonella foenum-graecum L.</i>
	<i>Momordica cochinchinensis (Lour.) Spreng.</i>	<i>Viburnum opulus L.</i>
	<i>Morinda citrifolia</i>	<i>Xenostegia tridentata</i>
11	Seeds	
	<i>Alcea pallida (Willd.) Waldst & Kit.</i>	<i>Nigella sativa L.</i>
	<i>Celosia argentea L.</i>	<i>Phaseolus vulgaris L.</i>
	<i>Ensete superbum (Roxb.) Cheesman</i>	<i>Piper nigrum</i>
	<i>Holarrhena pubescens</i>	<i>Pongamia pinnata</i>
	<i>Lupinus varius subsp.varius</i>	<i>Raphanus sativus L.</i>
	<i>Macrotyloma uniflorum (Lam.) Verdc.</i>	<i>Trachyspermum ammi</i>
	<i>Manilkara zapota (L.) P. Royen (kernel)</i>	<i>Trionella foenum-graecum L.</i>
	<i>Momordica cochinchinensis (Lour.) Spreng.</i>	<i>Vigna unguiculata (L.) Walp.</i>
12	Whole plant	
	<i>Achillea millefolium L.</i>	<i>Gallium verum L.</i>
	<i>Achyranthes aspera</i>	<i>Haloxylon stocksii (Boiss.) Benth. & Hook. f.</i>
	<i>Acmella oleracea</i>	<i>Heliotropium crispum Desf.</i>
	<i>Actinodaphne angustifolia Nees.</i>	<i>Herniaria hirsute L.</i>
	<i>Aeonium canariense (L.) Webb & Berthel</i>	<i>Hypericum montbretii Spach.</i>
	<i>Aerva lanata</i>	<i>Kalimeris indica (L.) Sch. Bip.</i>
	<i>Ajuga chamaepitys (L.) Schreb.</i>	<i>Lamium album L.</i>
	<i>Ammi majus L.</i>	<i>Lemanea fluviatilis L.</i>
	<i>Alcea flavovirens (Boiss.) Boiss</i>	<i>Lindernia ruelliooides (Colsm.) Pennell.</i>
	<i>Alisma plantago-aquatica L.</i>	<i>Ludwigia perennis L.</i>
	<i>Alternanthere tenella colla</i>	<i>Malvella sherardiana (L.) Jaub. & Spach.</i>
	<i>Amaranthus spinosus L.</i>	<i>Meiogyne minuta (G.Forst.)Less</i>
	<i>Amaranthus viridis</i>	<i>Mentha pulegium L.</i>
	<i>Bonnaya brachiate</i>	<i>Merremia emarginata</i>
	<i>Capsella bursa-pastoris Medik</i>	<i>Micromeria biflora</i>
	<i>Cassia italica (Mill.) Spreng.</i>	<i>Mukia maderaspatana</i>
	<i>Centella asiatica</i>	<i>Myriogyne minuta (G.Forst.) Less.</i>
	<i>Cerasus avium</i>	<i>Ocimum basilicum</i>
	<i>Ceterach aureum Buch.</i>	<i>Ocimum tenuiflorum</i>
	<i>Coccinia indica Wight & Arn.</i>	<i>Oldenlandia herbacea</i>
	<i>Costus spicatus (Jacq.) Sw.</i>	<i>Origanum majorana L.</i>
	<i>Citrullus colocynthis (L.) Shrad.</i>	<i>Orthosiphon grandiflorus Bold.</i>
	<i>Dendrophthoe falcata (L.f.) Ettingsh.</i>	<i>Orthosiphon stamineus Benth</i>
	<i>Desmodium microphyllum (Thunb.) DC</i>	<i>Parmelia perlata</i>
	<i>Desmodium styracifolium(Osbeck) Merr.</i>	<i>Pedalium murex L.</i>
	<i>Didymocarpus tomentosus Wight</i>	<i>Pergularia daemia</i>
	<i>Duchesnea indica (Jacks.) Focke.</i>	<i>Phyllanthus fraternus G.L. Webster</i>

S.No.	Class (Root)	Antiurolithiatic plants
	<i>Echinops echinatus</i> Roxb.	<i>Phyllanthus niruri</i>
	<i>Ehydra fluctuans</i> Lour.	<i>Phyllanthus urinaria</i>
	<i>Enhydra fluctuans</i> Lour.	<i>Pistacia lentiscus</i>
	<i>Erigeron karvinskianus</i> DC.	<i>Plantago coronopus</i>
	<i>Equisetum arvense</i>	<i>Polygonum aviculare</i>
	<i>Equisetum bogotense</i>	<i>Pratia nummularia</i> (Lam.) A.Braun & Asch.
	<i>Equisetum debile</i>	<i>Primula veris</i> L.
	<i>Euphorbia hirta</i> .	<i>Rubia cordifolia</i> L.
	<i>Euphorbia prostrata</i> Aiton	<i>Solanum nigrum</i> L.
	<i>Euphorbia serpens</i> Kunth.	<i>Sphaeranthus indicus</i> L.
	<i>Forsskaolea angustifolia</i> Retz.	<i>Teucrium chamaedrys</i> L.
	<i>Fragaria nilgerrensis</i>	<i>Teucrium scordium</i> L.
	<i>Fumaria officinalis</i> L.	<i>Withania somnifera</i> (L.) Dunal.

Table 3. Animal source with antiurolithiatic and diurtic activity

S.No.	Name	Parts used
1.	Burnt Scorpion	Whole part
2.	Sparrow	Brain
3.	Purified Earth worm	Whole part
4.	Burnt rabbit	Whole part

Table 4. Marine source with antiurolithiatic and diuretic activity

S.No.	Chemical constituent	Type of organism	Reference
1.	Chitosan (chitin) biopolymer	Marine and terrestrial organisms	Moacir Fernandes
2.	Fucoxanthine (active) carotenoid	Macroalgae, microalgae	Queiroz et al., [19]
3.	C-Phycocyanin	Macroalgae, microalgae Spirulina plantensis(blue-green algae)	Rulin Wang et al., [20] DéboraPez Jaeschke et al., [21]

4.4 Microbial Source

Oxalate degrading bacteria such as *Bifidobacterium* species, *bacillus* species, *oxalobacter formigenes*, *porphyromonas gingivalis*, *bacillus subtilis* and *Lactobacillus plantarum* show antiurolithiatic activity by degrading oxalate crystals [22].

Table 5. Mineral source with litholytic and diuretic activity

S.No.	Name
1.	Alum
2.	Burnt crystal
3.	Fish stones
4.	Lapis Judaicus / Jews Stone
5.	Potassium carbonate
6.	Potassium nitrate

4.5 Mineral Source

The mineral source with diuretic and litholytic activity are listed in Table 5 [1].

5. CONCLUSION

Urolithiasis is outstanding considerable malady of the urogenital system and also a great origin of morbidity [23-43]. With its several aetiologoun and high incidence of re occurrence, renal tract stone malady come up with a medical dispute [44-51].

Despite of many traditional curatives feasible for the urolithiasis, but re occurrence of stone formation still remains as a problem still to date there is no standard drug available and foremost disadvantage in the maturing of a standard drug may be the multifactorial kind of the disease and several chemical varieties of renal stones.

Many curatives has been serviced during the ages to cure renal stones. In the classic system of medicine, utmost of the therapies was taken from plants and they were proven to be useful though the rationale at the rear of their used is not Triumphant through systematic pharmacological and clinical studies apart from some compounded herbaceous medicines and plants. Herbal medicines contain several

phytonutrients and exert their beneficial outcomes by various mechanisms in treatment of urolithiasis.

DISCLAIMER

The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

CONSENT

It is not applicable

ETHICAL APPROVAL

It is not applicable

ACKNOWLEDGEMENTS

The authors would like to thank Sri Padmavati Mahila Visvavidyalayam, Tirupati. Collaborative research work and funded by University Grants Commission (UGC) (NFPwD) Scheme (Ref. No.: NFPwD-2018-2020-AND-8336) by the department of empowerment of persons with disabilities (Divyangjan), India for funding provided. My special thanks to Dr Y. Indira Muzib.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Mohammad Shamim Khan, Qamrul Hasan Lari, Mahmood Ahmad Khan. ANTI-UROLITHIATIC UNANI DRUGS – A REVIEW; World Journal of Pharmaceutical Research. 2016;5(12):279-294.
2. Deepti Sharma, Yadu Nandan Dey, Indu Sikarwar, Richa Sijoria, Manish M. Wanjari, Ankush D. Jadhav, In vitro study of aqueous leaf extract of *Chenopodium album* for inhibition of calcium oxalate and brushite crystallization, Egyptian Journal of Basic and Applied Sciences. 2016;3(22):164-171.
3. Dulce María González Mosquera, Yannarys Hernández Ortega, Pedro César Quero, Rafael Sosa Martínez, Luc Pieters, Antiurolithiatic activity of *Boldoa purpurascens* aqueous extract: An in vitro and in vivo study, Journal of Ethnopharmacology. 2020;253:112691. DOI: 10.1016/j.jep.2020.112691 Available:<https://doi.org/10.1016/j.ejbas.2016.02.001>
4. Humaira Bano, Nasreen Jahan, Shaikh Ajij Ahmed Makbul, Kumar BN, Sadique Husain, Atiya Sayed, Effect of *Piper cubeba* L. fruit on ethylene glycol and ammonium chloride induced urolithiasis in male Sprague Dawley rats, Integrative Medicine Research. 2018;7(4):358-365. DOI: 10.1016/j.imr.2018.06.005
5. Iman S, Saleem U and Ahmad B, Preclinical assessment of antiurolithiatic activity of *mangifera indica* seeds on ethylene glycol induced urolithiasis in rat model. Pak Vet J. 2021;41(1):92-96. DOI: 10.29261/pakvetj/2020.028
6. Jagannath N, Chikkannasetty SS, Govindadas D, Devasankaraiah G. Study of antiurolithiatic activity of *Asparagus racemosus* on albino rats. Indian J Pharmacol. 2012;44(5):576-579. DOI: 10.4103/0253-7613.100378
7. K V S R G Prasad, R Abraham, K Bharathi, K K Srinivasan. Evaluation of *Homonia riparia* Lour. for Antiurolithiatic Activity in Albino Rats, International Journal of Pharmacognosy, 1997;35(4): 278-283. DOI: 10.1076/phbi.35.4.278.13309
8. Ma Therese H Garcia, Carelle Juliene A Malaguit, Ashley Joy D Verte, Renz P Cometa, Maricel E Dimayacyac, et al. Antiurolithiatic activity of ethanolic extract of *Eleusine indica* Linn. (Poaceae) leaves in ethylene glycol-induced urolithiasis in male Sprague Dawley rats, Asia Pacific Journal of Allied Health Sciences. 2018;1. ISSN 2704-3568.
9. Mert İlhan, Burçin Ergene, İpek Süntar, Serkan Özbilgin, Gülcin Sultan Çitoğlu, M. Ayşe Demirel, et al. Preclinical Evaluation of Antiurolithiatic Activity of *Viburnum opulus* L. on Sodium Oxalate-Induced Urolithiasis Rat Model, Evidence-Based Complementary and Alternative Medicine. 2014;ArticleID 578103:10. Available:<https://doi.org/10.1155/2014/578103>

10. Mittal S, Tandon SK, Singla C Tandon. *In vitro* studies reveal antiurolithic effect of Terminalia arjuna using quantitative morphological information from computerized microscopy, International Brazilian Journal of Urology. 2015;41(5):935-944.
DOI: 10.1590/S1677-5538.IBJU.2014.0547
11. Mansoor M, Jamil M, Latif N, Muhammad S, Gull J, Shoab M, Khan A. Review Importance of Herbal Plants in the Management of Urolithiasis. Biological Sciences – PJSIR. 2019;62(1):61-66.
Available:<https://doi.org/10.52763/PJSIR.BIOL.SCI.62.1.2019.61.66>
12. Ram Prakash, Arunachalam, Narayanasamy, Prevalence and socio-demographic status on kidney stone patients in Thanjavur district, Tamil Nadu, India; Int J Community Med Public Health; 2019;6(5):1943-1947.
DOI: <http://dx.doi.org/10.18203/2394-6040.ijcmph20191614>
13. Shruti Gupta, Shamsher SK. Kidney stones: Mechanism of formation, pathogenesis and possible treatments; J Biomol Biochem. 2018;2(1):1-5.
14. Victoriano Romero, Haluk Akpinar, Dean G Assimos, Kidney stones: A global picture of prevalence, incidence, and associated risk factors; Rev Urol.; Spring. 2010;12(2-3):e86-96.
PMCID: PMC2931286.
15. Yu Liu, Yuntian Chen, Banghua Liao, Deyi Luo, Kunjie Wang, Hong Li. Guohua Zeng; Epidemiology of urolithiasis in Asia; Asian J Urol. 2018 5(4):205–214.
Available:<https://doi.org/10.1016/j.ajur.2018.08.007>
16. Vishal N Ratkalkar, Jack G Kleinman, Mechanisms of Stone Formation; Clin Rev Bone Miner Metab. 2011;9(3-4):187–197.
DOI: 10.1007/s12018-011-9104-8
17. Poonam Verma, Vinod Gauttam, Ajudhia N Kalia. Comparative pharmacognosy of Pashanbhed; Ayurveda Integr Med. 2014 Apr;5(2):104-8.
DOI: 10.4103/0975-9476.131728
18. Salman Ahmed, Muhammad Mohtasheemul Hasan, Zafar Alam Mahmood; Antiurolithiatic plants in different countries and cultures; JPP. 2016;5(1): 102-115.
19. Moacir Fernandes Queiroz , Karoline Rachel Teodosio Melo , Diego Araujo Sabry , Guilherme Lanzi Sassaki , Hugo Alexandre Oliveira Rocha Does the use of chitosan contribute to oxalate kidney stone formation?; Marine Drugs. 2014;13(1):141-58.
DOI: 10.3390/md13010141
20. Rulin Wang, Elsayed M Younis, Vishnu Priya Veeraraghavan, Chenfei Tian. Antiurolithiatic effect of Fucoxanthin on ethylene glycol-induced renal calculus in experimental rats. Journal of King Saud University – Science. 2020;32(3):1896-1901.
Available:<https://doi.org/10.1016/j.jksus.2020.01.027>
21. DéboraPez Jaeschke, Ingrid Rocha Teixeira, Ligia Damasceno Ferreira Marczak, GiovanaDomeneghini Mercali. Phycocyanin from Spirulina: A review of extraction methods and stability; Food Research International. 2021;143:110314.
DOI: 10.1016/j.foodres.2021.110314
22. Rouhi Afkari, Mohammad Mehdi Feizabadi, Alireza Ansari-Moghadam, Tahereh Safari, Mohammad Bokaeian. Simultaneous use of oxalate-degrading bacteria and herbal extract to reduce the urinary oxalate in a rat model: A new strategy; Int Braz J Urol. 2019;45(6):1249-1259.
DOI: 10.1590/S1677-5538.IBJU.2019.0167
23. Shazib Faridi, Somarendra Singh K. Preliminary study of prevalence of urolithiasis in North-Eastern city of India; J Family Med Prim Care. 2020 Dec 31;9(12):5939-5943.
DOI: 10.4103/jfmpc.jfmpc_1522_20
24. Mosquera DMG, Ortega YH, Quero PC, Martínez RS, Pieters L. Antiurolithiatic activity of Boldoa purpurascens aqueous extract: An in vitro and in vivo study. J Ethnopharmacol. 2020 May 10;253: 112691.
DOI: 10.1016/j.jep.2020.112691
25. Rathod N.R, Dipak Biswas, Chitme H.R, Sanjeev Ratna, I.S. Muchandi, Ramesh Chandra, Anti-urolithiatic effects of Punica granatum in male rats, Journal of Ethnopharmacology. 2012;140(2):234-238.
Available:<https://doi.org/10.1016/j.jep.2012.01.003>
26. Nishi Saxena, Ameeta Aggarwal, Study of Antiurolithiatic activity of a formulated herbal suspension, Herba Polonica-agro. 2015;61(2).
DOI: <https://doi.org/10.1515/hepo-2015-0014>
27. Kachchhi NR, Parmar RK, Tirgar PR, Desai TR, Bhalodia PN, Evaluation of the

- antiurolithiatic activity of methanolic extract of celosia argentea roots in rats, Kachchhi NR. et al. / International Journal of Phytopharmacology. 2012;3(3):249-255.
28. Olayeriju OS, Crown OO, Elekofehinti OO, et al. Effect of moonseed vine (*Trichilia gilletii* Staner) on ethane-1,2-diol-induced urolithiasis and its renotoxicity in Wistar albino rats. Afr J Urol. 2020;26: 4. Available:<https://doi.org/10.1186/s12301-020-0018-x>
29. Patel PK, Patel MA, Saralai MG, Gandhi TR, Antiurolithiatic Effects of *Solanum xanthocarpum* Fruit Extract on Ethylene-Glycol-Induced Nephrolithiasis in Rats, Journal of Young Pharmacists. 2012;4(3):164-170. DOI: 10.4103/0975-1483.100022
30. Paula PO Salem, Nátilie B Vieira, Daniela A Garcia, Karen J Nicácio, Danielle F Dias, Ana CC de Paula, et al., Anti-urolithiatic and anti-inflammatory activities through a different mechanism of actions of *Cissus gongyloides* corroborated its ethnopharmacological historic, Journal of Ethnopharmacology. 2020;253:112655. DOI: 10.1016/j.jep.2020.112655
31. Poppy Das, Kiran Kumar, Arunai Nambiraj, Reshma Rajan, Rajendra Awasthi, Kamal Dua, et al., Potential therapeutic activity of *Phlogacanthus thrysiformis* Hardow (Mabb) flower extract and its biofabricated silver nanoparticles against chemically induced urolithiasis in male Wistar rats, International Journal of Biological Macromolecules. 2017;103:621-629. DOI: 10.1016/j.ijbiomac.2017.05.096
32. R Vargas S, R.M Perez G, S Perez G, M.A Zavala S, C Perez G, Antiurolithiatic activity of *Raphanus sativus* aqueous extract on rats, Journal of Ethnopharmacology. 1999;68(1-3):335-338. Available:[https://doi.org/10.1016/S0378-8741\(99\)00105-1](https://doi.org/10.1016/S0378-8741(99)00105-1)
33. Ravinda V Karadi, Navneet B Gadge, Alagawadi KR, Rudraprabhu V Savadi. Effect of *Moringa oleifera* Lam. root-wood on ethylene glycol induced urolithiasis in rats, Journal of Ethnopharmacology; 2006;105(1-2):306-311. DOI: 10.1016/j.jep.2005.11.004
34. Sachin B Agawane, Vidya S Gupta, Mahesh J Kulkarni, Asish K Bhattacharya, Santosh S Koratkar, Vamkudoth Koteswara Rao, Patho-physiological evaluation of *Duranta erecta* for the treatment of urolithiasis, Journal of Ayurveda and Integrative Medicine. 2019; 10(1):4-11. DOI: 10.1016/j.jaim.2017.08.001
35. Sahithi Adepu, Raviraj Anand Devkar, Shilpee Chaudhary, Manganahalli Manjunath Setty, WITHDRAWN: Antiurolithiatic activity of *Alternanthera tenella* Colla in ethylene glycol induced urolithiasis in rats, Asian Journal of Pharmaceutical Sciences; 2015. ISSN 1818-0876. Available:<https://doi.org/10.1016/j.ajps.2015.11.003>
36. Tanzeer Kaur, Rakesh K. Bijarnia, Surinder K. Singla, Chanderdeep Tandon, *In vivo* efficacy of *Trachyspermum ammi* anticalcifying protein in urolithiatic rat model, Journal of Ethnopharmacology. 2009;126(3):459-462.
37. Tushar Shelke, Sandesh Wayal, Shiv Gunjegaokar, Sachin Gaikwad, Amol Shirasath, Sunil Hadke; An Overview on Indian Medicinal Plants with Antiurolithiatic Activity; Journal of Pharmaceutical Research & Clinical Practice. 2014;4(3): 33-40.
38. Sasikala V, Ramu Radha S, Bavaniamma Vijayakumari, *In vitro* evaluation of *Rotula aquatica* Lour. for antiurolithiatic activity, Journal of Pharmacy Research. 2013;6(3):378-382. Available:<https://doi.org/10.1016/j.jopr.2013.02.026>
39. Salman Ahmed, Muhammad Mohtasheemul Hasan, Zafar Alam Mahmood; Antiurolithiatic plants: Multidimensional pharmacology; JPP. 2016;5(2):04-24.
40. Salman Ahmed, Muhammad Mohtasheemul Hasan, Zafar Alam Mahmood. Antiurolithiatic plants: Formulations used in different countries and cultures; Pak. J. Pharm. Sci. 2016; 29(6):2129-2139.
41. Satish S, Mahesh CM, Gowda KPS, David Banji. Study on the Antiurolithiatic activity of *Cynodon dactylon* root stalk extract in Albino rats; CAB Direct, Biomed. 2009;4(4):384-391.
42. Shengbao Wang, Xiaoran Li, Junsheng Bao and Siyu Chen Protective potential of *Angelica sinensis* polysaccharide extract against

- ethylene glycol-induced calcium oxalate urolithiasis, Renal Failure. 2018;40(1):618-627.
43. Rahul Deo Yadav, Jain S.K, Shashi Alok, Alok Mahor, Jay Prakash Bharati, Manoj Jaiswal: Herbal plants used in the treatment of urolithiasis: IJPSR. 2011; 2(6):1412-1420.
44. Ajij A, Abdul W, Nasreen J, Alia Bilal, Syeda H, Efficacy of *Adiantum capillus veneris* Linn in chemically induced urolithiasis in rats, Journal of Ethnopharmacology. 2013;146(1):411-416. DOI: 10.1016/j.jep.2013.01.011
45. Atul M, Vishavas R, Dishant Desai, Jaymin M, Vivek Parekh, Evaluation for the anti-urolithiatic activity of *Launaea procumbens* against ethylene glycol-induced renal calculi in rats, Toxicology Reports. 2014; 1(2014):46-52. DOI: 10.1016/j.toxrep.2014.03.006
46. Kaleeswaran B, Ramadevi S, Murugesan R, Srigopalram.S, Suman.T, Balasubramanian.T, Evaluation of anti-urolithiatic potential of ethyl acetate extract of *Pedalium murex* L. on struvite crystal (kidney stone), Journal of Traditional and Complementary Medicine. 2017;9(1):24-37.
Available:<https://dx.doi.org/10.1016%2Fj.jcm.2017.08.003>
47. Kumar BN, Abdul Wadud, Nasreen Jahan, Ghulamuddin Sofi, Humaira Bano, Shaikh Ajij Ahmed Makbul, Sadique Husain, Antilithiatic effect of *Peucedanum grande* C. B. Clarke in chemically induced urolithiasis in rats, Journal of Ethnopharmacology. 2016;194(2016): 1122-1129. DOI: 10.1016/j.jep.2016.10.081
48. Vyas BA, Vyas RB, Joshi SV, Santani DD, Antiurolithiatic Activity of Whole-Plant Hydroalcoholic Extract of *Pergularia daemia* in Rats, Journal of Young Pharmacists. 2011;3(1):36-40. DOI: 10.4103/0975-1483.76417
49. Basavaraj M Dinnimath, Sunil S Jalalpure, Umesh K Patil, Antiurolithiatic activity of natural constituents isolated from *Aerva lanata*, Journal of Ayurveda and Integrative Medicine. 2017;8(4):226-232. DOI: 10.1016/j.jaim.2016.11.006
50. Bawari S, Sah AN, Tewari D. Anticalcifying effect of *Daucus carota* in experimental urolithiasis in Wistar rats. J Ayurveda Integr Med. 2020 Jul-Sep;11(3):308-315. DOI: 10.1016/j.jaim.2018.12.003
51. Adegbaju OD, Otunola GA, Afolayan AJ, Effects of growth stage and seasons on the phytochemical content and antioxidant activities of crude extracts of *Celosia argentea* L., Heliyon. 2020;6(6). DOI: 10.1016/j.heliyon.2020.e04086

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